Interactions between secondary structures and small molecules

Maria Grazia Nolli^{1*}, Andrea Patrizia Falanga¹, Maria Marzano¹, Alessandro D'Urso², Stefano D'Errico¹, Gennaro Piccialli¹, Giorgia Oliviero³ and Nicola Borbone¹.

1 - Department of Pharmacy, Università degli studi di Napoli Federico II, via D. Montesano 49 - 80131 Naples

2 - Department of Chemical sciences, Cittadella Universitaria, V.le A. Doria 6, 95125 Catania.

3 - Department of Molecular Medicines and Medical Biotechnologies, Università degli studi di Napoli Federico II, via D. Montesano 49 - 80131 Naples

E-mail: mariagrazia.nolli@unina.it

Oligonucleotide analogues (ODNs) are biomolecules with great scientific potential because of their remarkable properties (higher bioavailability, affinity with the target, stability, and resistance against nuclease degradation). Therefore, they can be used as nanoprobes and biosensors and to develop new materials in the nanotechnology field¹. ODNs and their analogues can assume several secondary structures which play an important role in Biology, Biotechnology, and Nanotechnology². In particular, ODNs rich in guanines can adopt secondary structures named G-quadruplexes, which are also very interesting from a diagnostic and therapeutic point of view³. In the human genome, there are G-quadruplex structures afferent to regions of genes of high regulatory importance (such as enhancers, promoters, and oncogenes). It has been shown that the stabilization and destabilization of these secondary structures can influence the onset of diseases. There is a field of research investigating the interactions between small molecules and DNA secondary structures, including Gquadruplexes. To study the interaction between G-Quadruplexes and ligands, in this paper, we focused our attention on the interactions between G-quadruplexes and modified Porphyrins, a class of macrocyclic compounds that play a significant role in the metabolism of living organisms⁴.

Bibliography

- 1. © 2013 Landes Bioscience SpeciAL FocuS ReVieW SpeciAL FocuS ReVieW; 2013. www.landesbioscience.com.
- 2. SantaLucia, J.; Hicks, D. The Thermodynamics of DNA Structural Motifs. Annual Review of Biophysics and Biomolecular Structure. 2004, pp 415–440. https://di.org/10.1146/annurev.biophys.32.110601.141800.
- Oliviero, G.; D'Errico, S.; Pinto, B.; Nici, F.; Dardano, P.; Rea, I.; De Stefano, L.; Mayol, L.; Piccialli, G.; Borbone, N. Self-Assembly of g-Rich Oligonucleotides Incorporating a 3'-3' Inversion of Polarity Site: A New Route towards G-Wire DNA Nanostructures. ChemistryOpen 2017, 6 (4), 599–605. https://doi.org/10.1002/open.201700024.
- 4. Biesaga, M.; Pyrzyn´ska, K.; Pyrzyn´ska, P.; Trojanowicz, M. Porphyrins in Analytical Chemistry. A Review; 2000; Vol. 51. www.elsevier.com/locate/talanta.