



# The Diet–Multiple Sclerosis Connection: Oxidative Stress and Emerging Mechanisms

Candida Bucciero<sup>1</sup> · Alessandra Croce<sup>1</sup> · Giuliano Castellano<sup>1</sup> · Francesco Beguinot<sup>1</sup> · Pietro Formisano<sup>1</sup> · Giuseppe Portella<sup>1</sup> · Luca Ulianich<sup>2</sup> · Francesca Fiory<sup>1</sup> · Anna Maria Malfitano<sup>1</sup>

Received: 18 December 2025 / Accepted: 10 February 2026  
© The Author(s) 2026

## Abstract

Multiple sclerosis (MS) is an autoimmune neuroinflammatory disease resulting in myelin degeneration and progressive disability. Oxidative stress plays a crucial role in MS pathogenesis and progression. Nuclear factor erythroid 2-related factor 2 (Nrf2) is a master regulator of the antioxidant mechanisms and its upregulation is associated with beneficial effects in MS. Among the environmental factors influencing MS onset and progression, diet represents a promising non-pharmacological strategy to modulate Nrf2, potentially improving MS outcomes. Indeed, several natural compounds present in Mediterranean, ketogenic and Paleolithic diets can enhance Nrf2 activity, and exert beneficial effects in preclinical models of MS. In this review, we summarize the key role of oxidative stress in MS and highlight how dietary regimens and Nrf2-modulating natural compounds might have therapeutic potential for MS patients. Additionally, we discuss emerging and still poorly explored mechanisms beyond classical Nrf2 activation, including epigenetic regulation and the stability of DNA/RNA secondary structures known as G-quadruplexes, which are involved in gene expression regulation and may represent novel nutrition-based therapeutic targets. However, while Nrf2 modulation by diet is supported by preclinical and limited clinical evidence, targeting G-quadruplexes as a strategy to counteract oxidative stress in MS remains largely speculative and requires further investigation. Notably, epigenetic mechanisms and G-quadruplexes may represent innovative targets of Nrf2-boosting dietary natural compounds for the development of supplemental therapeutic strategies for MS.

**Keywords** Multiple sclerosis · Oxidative stress · Diets · Epigenome · G-quadruplexes

## Introduction

Multiple sclerosis (MS) is an immune-mediated, multifactorial chronic demyelinating disease of the central nervous system (CNS), characterized by neurodegeneration, inflammation and axonal loss [1]. The resulting muscle spasticity

and weakness lead to motor impairments and a compromised quality of life in MS patients [2]. The pathogenic mechanism involved in MS is characterized by sustained infiltration of autoreactive T cells across the blood–brain barrier into the CNS [3, 4], driving myelin sheath and axon destruction [2].

Oxidative stress (OS) results from excessive formation of reactive oxygen species (ROS), mitochondrial dysfunction and a poor or compromised antioxidant defense system [1]. OS plays a key role in MS, sustaining both the inflammatory and neurodegenerative process of the disease [5]. In the acute phase, OS contributes to the initiation of inflammation, whereas in the chronic phase it sustains neurodegeneration [6].

Intriguingly, plasma isolated from MS patients shows increased levels of OS markers together with altered antioxidant defenses [7]. Autoreactive T lymphocytes induce microglia to release large amounts of ROS, cytokines, oxidative products, and free radicals [8]. Mitochondrial damage occurs at early stages of MS and plays a key role in

---

Francesca Fiory and Anna Maria Malfitano contributed equally as last authors.

✉ Francesca Fiory  
francesca.fiory@unina.it

✉ Anna Maria Malfitano  
annamaria.malfitano@unina.it

<sup>1</sup> Department of Translational Medical Sciences, University of Naples Federico II, 80131 Naples, Italy

<sup>2</sup> Institute of Endotypes in Oncology, Metabolism and Immunology “G. Salvatore”—National Research Council (IEOMI-CNR), 80131 Naples, Italy

inflammation and disease progression [9] by increasing the production of toxic ROS, triggering apoptosis, demyelination and neurodegeneration [10]. OS further impairs ATP transport along axons and compromises cellular homeostasis, particularly in neurons and oligodendrocytes, which are notably vulnerable due to weaker antioxidant defenses.

Nuclear factor erythroid 2-related factor 2 (Nrf2), a transcription factor regulating antioxidant gene expression, has emerged as a key modulator of OS in MS. Preclinical studies suggest that Nrf2 activation reduces inflammatory damage and promotes neuronal survival [11, 12]. However, heterogeneity among experimental models and the scarcity of human studies currently limit clinical translation. Interestingly, several dietary regimens, such as Mediterranean, ketogenic, and Paleolithic diets, have been proposed to modulate OS via Nrf2 activation. Despite promising preclinical evidence, clinical outcomes remain inconsistent, as differences in disease stage, patient genetics, and adherence strongly influence efficacy.

This review summarizes the role of OS and Nrf2 in MS and provides a critical synthesis of recent literature on the relevance of diet in MS and the ability of dietary natural compounds in Nrf2 modulation, highlighting updated clinical insights, mechanistic gaps, and translational challenges. Finally, we discuss emerging mechanisms potentially involved in Nrf2 regulation, including epigenetic regulation and the stability of DNA/RNA G-quadruplex structures, which may represent novel nutritional targets to improve the efficacy of current MS therapies.

## Relevance of Nrf2 in MS

Nrf2 orchestrates the cellular antioxidant response through the regulation of genes encoding detoxifying enzymes such as heme oxygenase (HO)-1, NAD(P)H:quinone oxidoreductase 1 (NQO-1), and glutathione S-transferases. Upon exposure to ROS, Nrf2 dissociates from its negative regulator Kelch-like ECH-associated protein 1 (Keap1) and translocates to the nucleus, where it binds to the antioxidant response element (ARE) in the promoter region of these and other genes involved in antioxidant defense and detoxification [13].

Interestingly, several chronic conditions such as respiratory, cardiovascular, kidney, liver and neurodegenerative diseases exhibit an imbalanced Nrf2 pathway [14]. For instance, in Parkinson's disease (PD), Nrf2 downregulation induces  $\alpha$ -synuclein aggregation [15], while Nrf2 activation protects dopaminergic neurons from OS-induced neurotoxicity [16]. In Alzheimer's disease (AD), the decrease of Nrf2 levels impairs autophagy and leads to the accumulation of amyloid- $\beta$  and tau [17]. The central role of Nrf2 in these pathologies has prompted investigations aimed at enhancing Nrf2 activity to improve cellular oxidative defense.

In MS, impaired Nrf2 activation is critical for preventing mitochondrial failure, OS, neuroinflammation, and neurodegeneration [18]. Dysfunctions in the Nrf2 pathway disrupt redox homeostasis, leading to increased ROS production [19] and the activation of other redox-sensitive transcription factors, such as the pro-inflammatory nuclear factor kappa-light chain-enhancer of activated B cells (NF- $\kappa$ B) and activator protein 1 (AP-1). Consequently, NF- $\kappa$ B induces the expression of genes involved in MS pathogenesis, such as inducible nitric oxide synthase (iNOS), interleukin 1 $\alpha$ / $\beta$  (IL-1 $\alpha$ / $\beta$ ), tumor necrosis factor  $\alpha$  (TNF- $\alpha$ ), and several growth factors [20]. Additionally, the downstream effectors of Nrf2, such as HO-1, play anti-inflammatory roles in experimental autoimmune encephalomyelitis (EAE) and in MS [21]. In peripheral blood mononuclear cells from MS patients, decreased HO-1 expression correlates with disease exacerbation [22]. However, in autopsy specimens of brain samples and spinal cord from MS patients, Nrf2, HO-1 and NQO-1 were upregulated both within and around active lesions [23], suggesting a role in the endogenous antioxidant defense. Moreover, Nrf2 expression seems to be cell-type-specific [24]: increased levels were observed in macrophages and astrocytes within active lesions [25], and in oligodendrocytes at the lesion's edges [26]. Notably, damaged oligodendrocytes in EAE lesions exhibit relatively low levels of Nrf2, suggesting their high vulnerability to OS [12, 27]. Consistently, Nrf2-deficient EAE mice display a more rapid disease onset and exacerbated clinical severity, accompanied by enhanced lesions, infiltrating immune cells, higher microglial activation, and visual dysfunction. In contrast, Nrf2 activation correlates with reduced demyelination and neuroinflammation in EAE and other preclinical MS models [28, 29].

Collectively, these studies suggest that strategies to enhance endogenous Nrf2 activation may protect the CNS from oxidative damage. The FDA-approval of dimethyl fumarate (DMF, Tecfidera) [30, 31] as an Nrf2-modulating drug supports the therapeutic relevance of Nrf2 in MS. Upon Nrf2 activation, DMF upregulates the downstream target genes such as NQO1 and HO-1 [32] and induces a shift toward peripheral regulatory immune cells [33], ultimately leading to improved clinical outcomes. Overall, targeting Nrf2 represents a feasible neuroprotective strategy in MS.

## Influence of Diets on Nrf2 Activation

The etiology of MS involves multiple factors including genetic, epigenetic, and environmental determinants. Among the environmental factors, diet is easily modifiable and is known to affect MS progression and symptoms [34]. Indeed, several dietary regimens have been reported to exert beneficial effects in MS as summarized in Table 1.

**Table 1** Dietary regimens beneficial for MS

|                              | Effects in MS  | Mechanism/s                                       | References        |
|------------------------------|--|---|-------------------|
| KD/Fast mimicking diet       | Increase of lean mass; improved body composition; reduced depression and fatigue; axon remyelination | Anti-inflammatory; modification of gut microbiota | [93–97]           |
| Paleolithic diet/Swank diet  | Improved walking and cognitive performances, fatigue and quality of life                             | Improvement of serum fatty acids profile          | [44, 98, 101–105] |
| Anti-inflammatory regimen/MD | Improved fatigue, inflammation and clinical manifestations   | Anti-inflammatory                                 | [36, 106–109]     |

The table reports dietary regimens and their beneficial effects in MS along with the type of mechanism elicited

Interestingly, these diets can activate Nrf2, thereby contributing to the induction of antioxidant and anti-inflammatory defenses.

In particular, the Mediterranean diet (MD) and Okinawan diet, which is very similar to Paleolithic diet, are both effective in boosting Nrf2 signaling [35]. The MD is characterized by moderate consumption of fish and dairy products and low intake of meat, in favor of fruit, vegetables and whole grains [36]. The Okinawan diet, considered a modern adaptation of ancestral human diet, is characterized by a high intake of sweet potatoes, vegetables, and plant-based foods, together with a low consumption of meat [35]. In contrast, the Paleolithic diet favors vegetables, fruits, lean meats, fish, and eggs while excluding grains, dairy products, added sugars, and processed foods [37]. The ketogenic diet (KD) has been shown to reduce OS and inflammation by activating Nrf2 in murine models of spinal cord injury [38]. Studies in mice showed that KD initially modifies hippocampal mitochondrial H<sub>2</sub>O<sub>2</sub> production, which activates the Nrf2 pathway in brain and liver, leading to the induction of protective proteins and the amelioration of the mitochondrial redox state [39]. Similarly, intermittent fasting regimens suppress inflammatory responses by inducing the sirtuin (SIRT) 3/Nrf2/HO-1 pathway in murine models of intracerebral hemorrhage [40]. In addition, fast-mimicking diets, proposed as intermittent nutritional interventions are capable of reducing inflammation and OS by alternating an initial phase of overall caloric restriction with a subsequent phase of normal dietary intake [41, 42]. However, it should be noted that KD and Paleolithic diet may be associated with potential risks, such as nutrient deficiencies, dyslipidemia, and reduced fiber intake [43–45], highlighting the need for careful monitoring and individualized dietary planning in MS patients. These findings suggest a strong rationale for Nrf2-targeting diets; however, translation to humans remains limited due to differences between rodent models

and human pathology, variability in diet adherence, and stage-specific responsiveness in MS patients. Moreover, many available studies are limited in sample size, short-term, and lack robust control groups.

### Nrf2-Targeting Dietary Natural Compounds and Their Benefits in MS

Beyond whole dietary patterns associated with Nrf2 activation, recent research identified specific health-promoting natural dietary compounds able to upregulate Nrf2. These compounds belong to different classes of bioactive molecules, including phenolic antioxidants, isothiocyanates from cruciferous vegetables, terpenoids and alkaloids. Several natural compounds, able to regulate Nrf2, have shown beneficial effect in animal models of EAE or demyelination and are currently considered potential dietary supplements for MS. Among these compounds, resveratrol, a natural polyphenol found in several plants, has been shown to activate Nrf2 signaling and promote the expression of antioxidant enzymes such as SOD and glutathione peroxidase, thereby contributing to free radical scavenging and reduction of OS [46]. In lung macrophages, resveratrol modulates Keap1–Nrf2 interaction by targeting the Ile28 residue in DLG motif of Nrf2, inducing conformational changes that alter KEAP1–DLG binding. The disruption of this interaction decreases Keap1-mediated proteasomal degradation of Nrf2, leading to increased Nrf2 expression and nuclear translocation [47]. A similar mechanism has been observed in the brain of AD mouse model, where resveratrol reduces the cytoplasmic Nrf2 levels, while enhancing its nuclear translation rate [48]. In a PD mouse model, resveratrol increases both Nrf2 and SIRT1 expression [49], in turn, SIRT1 activation may indirectly enhance Nrf2 stability via deacetylation-dependent mechanisms [50]. Additionally, in PC12 cells, resveratrol stimulates Nrf2-dependent transcription by activating the Phosphoinositide 3-kinase (PI3K)/Akt and Extracellular Signal-Regulated Kinase (ERK)1/2 pathways [51, 52].

Curcumin, another polyphenol derived from the rhizome of *Curcuma longa*, activates Keap1-Nrf2-ARE signaling pathway by modifying cysteine sulfhydryl groups in Keap1 [53–55]. Curcumin has also been shown to indirectly activate the ARE system by stimulating the upstream kinases Protein kinase C (PKC) and p38 Mitogen-Activated Protein Kinase (MAPK), which are required for full activation of HO-1, and by inhibiting protein phosphatase activity [56, 57]. Interestingly, two curcumin analogues have also been reported to increase Nrf2 and HO-1 protein expression and promote Nrf2 nuclear translocation, exerting antioxidant and neuroprotective effects in PC12 cells [58]. Notably, the therapeutic potential of curcumin in MS patients has been investigated in several clinical trials [59].

Similarly, several studies reported that the green tea-derived polyphenol epigallocatechin-3-gallate (EGCG) [60] activates the Nrf2 pathway in neuronal [61, 62] and microglial cells [63]. The underlying molecular mechanisms, including activation of ERK and PI3K, have been investigated in different cell types, such as human mammary epithelial cells [64] and hepatocytes [65].

Other natural compounds able to upregulate Nrf2 in the CNS, include matrine, a quinolizidine alkaloid derived from *Radix Sophorae Flave* [66]; withametelin, a phytosterol isolated from the leaves of *datura innoxia* plant [67]; piperine, the main bioactive alkaloid of black pepper, sulforaphane, an isothiocyanate found in cruciferous vegetables [68, 69]; myricetin, a flavonoid originally isolated from the bark of the tree *Myrica rubra* [70]; and many others [71]. Regarding the molecular mechanisms of Nrf2 activation, matrine exerts neuroprotective effects by upregulating Nrf2 via SIRT1 [72], withametelin enhances Nrf2 expression by decreasing Keap-1 levels in the CNS [67], and myricetin induces Nrf2 nuclear translocation in a murine cuprizone-induced demyelination model [73]. Interestingly, recent studies have highlighted the involvement of epigenetic changes in Nrf2 activation [74]. For example, sulforaphane increases Nrf2 expression and nuclear translocation while reducing DNA methylation of the Nrf2 promoter in a cellular model of AD [75]. However, the epigenetic modulation of Nrf2 by dietary compounds remains poorly investigated in the context of MS. Importantly, the beneficial effects of Nrf2-targeting natural compounds have been clearly demonstrated in animal models of demyelination. For instance, resveratrol is among the most extensively studied natural compounds and has shown promising neuroprotective and anti-inflammatory effects in EAE mice, where its supplementation reduced neuroinflammation and ameliorated clinical symptoms [46]. Curcumin also elicits effects comparable to resveratrol on Nrf2 signaling pathway and in MS-related experimental models [76]. Similarly, the Nrf2 upregulation induced by

piperine significantly ameliorates memory performance and myelin repair in a rat model of demyelination [77]. Finally, in the cuprizone-induced demyelination model, myricetin improves motor hyperactivity and behavioral deficits [73], and matrine ameliorates clinical signs in EAE [78].

### Potential Mechanisms Beyond Nrf2 Modulation: Epigenome and G-Quadruplexes

Beyond classical Nrf2 activation pathways, emerging evidence highlights that nutritional compounds may influence oxidative homeostasis through additional modulatory molecular mechanisms, including epigenetic regulation and the folding of DNA/RNA G-quadruplex secondary structures, which play key roles in gene expression. Several dietary supplements with antioxidant properties have been reported to act as epigenetic modulators [79]. Interestingly, Nrf2 expression can be influenced by epigenetic mechanisms, such as DNA methylation [74], histone modifications, and interactions with non-coding RNAs [80]. Recent studies have shown that targeting epigenetic modifications within the Nrf2 pathway represents a promising therapeutic strategy using dietary natural compounds such as curcumin, sulforaphane, and resveratrol [79], paving the way for the identification of additional dietary phytochemicals capable of modulating the human epigenome and enhancing antioxidative defense.

To date, the therapeutic potential of G-quadruplex folding modulation has been explored primarily in cancer, where stabilization of these motifs exerts important gene regulatory effects [81–83]. However, direct investigations of G-quadruplex structures in neurodegenerative diseases, including MS, are still at an early stage and warrant further study [84]. Nevertheless, G-quadruplex structures may theoretically affect the Nrf2 pathway through at least two molecular mechanisms. The first mechanism involves the modulation of autophagy, a process essential for the clearance of damaged cellular components and for remyelination following demyelinating injury in MS [85]. Autophagy has been shown to be regulated by G-quadruplexes in neurons [86] and contributes to redox balance by counteracting ROS accumulation via the sequestration and degradation of Keap1, thereby promoting release and activation of Nrf2 [87]. A second potential mechanism involves the presence of G-quadruplex-forming sequences in the promoter region of the Nrf2 gene [88]. Indeed, the 5' untranslated region (5'UTR) of Nrf2 mRNA can adopt G-quadruplex conformations that interact with elongation factor 1 alpha (EF1a), facilitating Nrf2 protein translation under conditions of OS and activating the cellular antioxidant response [89]. Importantly, several dietary natural compounds, including

resveratrol, curcumin, and piperine elicit antioxidant effects and can target G-quadruplex motifs [90–92]. In particular, polyphenols have recently been described as G-quadruplex stabilizers of interest for biomedical applications [93–98].

### Translational Limitations of Nrf2-Targeting Nutritional Interventions and Future Perspectives

Dietary natural compounds that boost Nrf2 show promising effects in preclinical MS models by targeting immune regulation, OS suppression, and supporting myelin regeneration. However, clinical trials testing the efficacy of Nrf2-boosting diets and natural compounds, summarized in Table 2, have reported variable outcomes in terms of fatigue amelioration, modulation of metabolic parameters, inflammatory biomarkers, or Magnetic Resonance Imaging (MRI) activity. Despite robust preclinical evidence, differences in study design, disease stage, nutrient dose, and follow-up duration, along with poor bioavailability and lack of standardized formulations, significantly limit clinical translation. For instance, the pronounced lipophilicity of curcumin and resveratrol reduces water solubility, intestinal absorption and systemic exposure [99], which may partly explain the modest or inconsistent outcomes reported in several clinical trials.

Nanoencapsulation has been proposed as a potential strategy to overcome these challenges by protecting natural compounds from degradation, oxidation, and pH extremes, while enhancing solubility, absorption and enabling controlled release. Nevertheless, current research remains focused on evaluating the safety profile of these delivery systems, particularly with respect to long-term exposure and potential toxicity [99]. Additionally, intrinsic differences between animal models and human MS, including disease heterogeneity, treatment duration, dosing regimens, and inter-individual variability, further contribute to the translational gap [100].

These limitations highlight the need for cautious interpretation of preclinical data and underscore the importance of well-designed clinical studies with standardized formulations, optimized delivery strategies, and comprehensive radiological evaluations to accurately assess the therapeutic potential of Nrf2-targeting nutritional interventions in MS.

Furthermore, additional studies are needed to uncover new molecular mechanisms involved in Nrf2 activation that may be targeted by dietary natural compounds. In particular, the potential link between dietary natural compounds, G-quadruplex stabilization, and Nrf2 activation in MS remains entirely hypothetical. To date, no direct experimental or clinical evidence supports a role for G-quadruplex-mediated antioxidant mechanisms in MS models or patients. Exploring these mechanisms may also help identify natural dietary compounds that specifically target G-quadruplexes, potentially paving the way for innovative nutrition-based therapeutic strategies in MS.

**Table 2** Clinical trials exploring effects of dietary interventions in MS

| Intervention (diet or nutrient)                             | Study design/Trial/Population and size   | Main reported outcome(s)  | References |
|---|--|---|------------|
| MD  | Randomized controlled trial, 1-year follow-up; 34 MS patients on Mediterranean-like diet vs 38 on standard healthy diet            | ↓ fatigue scores; no significant improvement in cognitive status  | [110]      |
| KD  | 18-month randomized controlled trial in 3 groups: 1) KD vs 2) standard diet vs 3) fasting diet in RRMS; 105 enrolled, 81 completed | Primary: no difference in new T2 MRI lesions among groups. Secondary/exploratory: in group 1 improvement in cognition at 18 months; in group 2 ↓ Neurofilament light chain (NFL) at 9 months; cardiometabolic markers improved in groups 1 and 3 and were partially associated with clinical outcomes | [111]      |
| Curcumin (as dietary supplement) add-on to standard therapy | Phase II randomized, placebo-controlled trial (with subcutaneous interferon $\beta$ -1 $\alpha$ ) in relapsing MS                  | At 12 months: lower proportion of patients with combined unique active (CUA) MRI lesions in curcumin group vs placebo; at 24 months no significant difference in new/enlarging T2 lesions, relapses, disability, or neurodegeneration markers   | [59]       |
| Resveratrol (500 mg/day)                                    | Double-blind randomized placebo-controlled trial, 8 weeks, 55 MS patients (RRMS)   | Significant reduction in inflammatory marker (TNF- $\alpha$ ) and oxidative stress marker like malondialdehyde (MDA) vs placebo; no change in fatigue score   | [112]      |

The table reports the type of intervention (diet or nutrient) adopted in the clinical trial, the cohort of patients recruited and the main outcomes

## Conclusions

It is well established that Nrf2 is a master regulator of cellular antioxidant defense. To date, numerous dietary natural compounds have been identified for their ability to activate Nrf2 and for their beneficial effects in supporting myelin integrity and reducing neuroinflammation in preclinical models of MS. In this review, we summarized experimental studies highlighting the relevance of the Nrf2 signaling pathway in MS and discussed the Nrf2-targeting properties of specific dietary regimens, including Mediterranean, ketogenic, and Paleolithic diets. We also focused on specific dietary natural compounds able to activate Nrf2 that are commonly consumed in daily diet and have demonstrated beneficial effects in MS-related experimental models. Interestingly, some of these compounds can modulate the epigenome and potentially influence G-quadruplex folding. Notably, while dietary modulation of Nrf2 is supported by experimental evidence, the involvement of G-quadruplex-mediated mechanisms in MS remains largely theoretical, with current insights derived primarily from other biological contexts. Therefore, mechanistic studies and well-designed clinical trials are required to validate these emerging targets and to optimize diet-based interventions as supplemental therapies for MS. To date, robust randomized controlled trials are still lacking, and major challenges related to target selectivity and poor bioavailability of dietary compounds remain unresolved. In conclusion, from a future perspective, combining conventional pharmacological approaches with targeted dietary strategies may represent a promising approach to reduce OS and improve both the efficacy and safety of current MS therapies.

**Author Contribution** C.B.: Literature search, data curation, original draft preparation A.C., G.C.: Literature search and data curation L.U., F.B., P.F., G.P.: review, editing and supervision F.F., A.M.M.: Writing – review, editing and supervision All authors have read and agreed to the published version of the manuscript.

**Funding** Open access funding provided by Università degli Studi di Napoli Federico II within the CRUI-CARE Agreement. This work was supported by: PROGETTI DI RICERCA DI RILEVANTE INTERESSE NAZIONALE (PRIN) 000013\_PRIN\_2022\_Malfitano\_A.M.\_cod.20224JSC3M.

**Data Availability** No datasets were generated or analysed during the current study.

## Declarations

**Competing interests** The authors declare no competing interests.

**Open Access** This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source,

provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit <http://creativecommons.org/licenses/by/4.0/>.

## References

- Ranieri R, Laezza C, Bifulco M, Marasco D, Malfitano AM (2016) Endocannabinoid system in neurological disorders. *Recent Pat CNS Drug Discov* 10(2):90–112. <https://doi.org/10.2174/1574889810999160719105433>
- Jakimovski D, Bittner S, Zivadnov R, Morrow SA, Benedict RH, Zipp F, Weinstock-Guttman B (2024) Multiple sclerosis. *Lancet* (London, England) 403(10422):183–202. [https://doi.org/10.1016/s0140-6736\(23\)01473-3](https://doi.org/10.1016/s0140-6736(23)01473-3)
- Compston A, Coles A (2008) Multiple sclerosis. *Lancet* (London, England) 372(9648):1502–1517. [https://doi.org/10.1016/s0140-6736\(08\)61620-7](https://doi.org/10.1016/s0140-6736(08)61620-7)
- Malfitano AM, Matarese G, Pisanti S, Grimaldi C, Laezza C, Bisogno T, Di Marzo V, Lechler RI et al (2006) Arvanil inhibits T lymphocyte activation and ameliorates autoimmune encephalomyelitis. *J Neuroimmunol* 171(1–2):110–119. <https://doi.org/10.1016/j.jneuroim.2005.09.005>
- Miller ED, Dziedzic A, Saluk-Bijak J, Bijak M (2019) A review of various antioxidant compounds and their potential utility as complementary therapy in multiple sclerosis. *Nutrients*. <https://doi.org/10.3390/nu11071528>
- Adamczyk B, Adamczyk-Sowa M (2016) New insights into the role of oxidative stress mechanisms in the pathophysiology and treatment of multiple sclerosis. *Oxid Med Cell Longev* 2016:1973834. <https://doi.org/10.1155/2016/1973834>
- Ferretti G, Bacchetti T, Principi F, Di Ludovico F, Viti B, Angelelli VA, Danni M, Provinciali L (2005) Increased levels of lipid hydroperoxides in plasma of patients with multiple sclerosis: a relationship with paraoxonase activity. *Mult Scler J* 11(6):677–682. <https://doi.org/10.1191/1352458505ms12400a>
- Lim JL, Wilhelmus MM, de Vries HE, Drukarch B, Hoozemans JJ, van Horssen J (2014) Antioxidative defense mechanisms controlled by Nrf2: state-of-the-art and clinical perspectives in neurodegenerative diseases. *Arch Toxicol* 88(10):1773–1786. <https://doi.org/10.1007/s00204-014-1338-z>
- Haider L (2015) Inflammation, iron, energy failure, and oxidative stress in the pathogenesis of multiple sclerosis. *Oxid Med Cell Longev* 2015:725370. <https://doi.org/10.1155/2015/725370>
- Tobore TO (2021) Oxidative/nitroxidative stress and multiple sclerosis. *J Mol Neurosci* 71(3):506–514. <https://doi.org/10.1007/s12031-020-01672-y>
- Tonev D, Momchilova A (2023) Oxidative stress and the nuclear factor erythroid 2-related factor 2 (Nrf2) pathway in multiple sclerosis: focus on certain exogenous and endogenous Nrf2 activators and therapeutic plasma exchange modulation. *Int J Mol Sci*. <https://doi.org/10.3390/ijms242417223>
- Johnson DA, Amirahmadi S, Ward C, Fabry Z, Johnson JA (2010) The absence of the pro-antioxidant transcription factor Nrf2 exacerbates experimental autoimmune encephalomyelitis. *Toxicol Sci* 114(2):237–246. <https://doi.org/10.1093/toxsci/kfp274>
- He F, Ru X, Wen T (2020) NRF2, a transcription factor for stress response and beyond. *Int J Mol Sci*. <https://doi.org/10.3390/ijms21134777>

14. Wang T, Liu M, Li X, Zhang S, Gu H, Wei X, Wang X, Xu Z et al (2024) Naturally-derived modulators of the Nrf2 pathway and their roles in the intervention of diseases. *Free Radic Biol Med* 225:560–580. <https://doi.org/10.1016/j.freeradbiomed.2024.09.035>
15. He Q, Song N, Jia F, Xu H, Yu X, Xie J, Jiang H (2013) Role of  $\alpha$ -synuclein aggregation and the nuclear factor E2-related factor 2/heme oxygenase-1 pathway in iron-induced neurotoxicity. *Int J Biochem Cell Biol* 45(6):1019–1030. <https://doi.org/10.1016/j.biocel.2013.02.012>
16. Yang XX, Yang R, Zhang F (2022) Role of Nrf2 in Parkinson's disease: toward new perspectives. *Front Pharmacol* 13:919233. <https://doi.org/10.3389/fphar.2022.919233>
17. De Plano LM, Calabrese G, Rizzo MG, Oddo S, Caccamo A (2023) The role of the transcription factor Nrf2 in Alzheimer's disease: therapeutic opportunities. *Biomolecules*. <https://doi.org/10.3390/biom13030549>
18. Gan L, Johnson JA (2014) Oxidative damage and the Nrf2-ARE pathway in neurodegenerative diseases. *Biochim Biophys Acta* 1842(8):1208–1218. <https://doi.org/10.1016/j.bbadis.2013.12.011>
19. Zgorzyska E, Dziedzic B, Walczewska A (2021) An overview of the Nrf2/ARE pathway and its role in neurodegenerative diseases. *Int J Mol Sci*. <https://doi.org/10.3390/ijms22179592>
20. Winyard PG, Blake DR (1997) Antioxidants, redox-regulated transcription factors, and inflammation. *Adv Pharmacol (San Diego, Calif)* 38:403–421. [https://doi.org/10.1016/s1054-3589\(08\)60993-x](https://doi.org/10.1016/s1054-3589(08)60993-x)
21. Schluesener HJ, Seid K (2000) Heme oxygenase-1 in lesions of rat experimental autoimmune encephalomyelitis and neuritis. *J Neuroimmunol* 110(1–2):114–120. [https://doi.org/10.1016/s0165-5728\(00\)00352-0](https://doi.org/10.1016/s0165-5728(00)00352-0)
22. Fagone P, Patti F, Mangano K, Mammana S, Coco M, Touil-Boukoffa C, Chikovani T, Di Marco R et al (2013) Heme oxygenase-1 expression in peripheral blood mononuclear cells correlates with disease activity in multiple sclerosis. *J Neuroimmunol* 261(1–2):82–86. <https://doi.org/10.1016/j.jneuroim.2013.04.013>
23. van Horssen J, Drexhage JA, Flor T, Gerritsen W, van der Valk P, de Vries HE (2010) Nrf2 and DJ1 are consistently upregulated in inflammatory multiple sclerosis lesions. *Free Radic Biol Med* 49(8):1283–1289. <https://doi.org/10.1016/j.freeradbiomed.2010.07.013>
24. Lee DH, Gold R, Linker RA (2012) Mechanisms of oxidative damage in multiple sclerosis and neurodegenerative diseases: therapeutic modulation via fumaric acid esters. *Int J Mol Sci* 13(9):11783–11803. <https://doi.org/10.3390/ijms130911783>
25. Cores Á, Carmona-Zafra N, Clerigú J, Villacampa M, Menéndez JC (2023) Quinones as neuroprotective agents. *Antioxidants*. <https://doi.org/10.3390/antiox12071464>
26. Licht-Mayer S, Wimmer I, Traffehn S, Metz I, Brück W, Bauer J, Bradl M, Lassmann H (2015) Cell type-specific Nrf2 expression in multiple sclerosis lesions. *Acta Neuropathol* 130(2):263–277. <https://doi.org/10.1007/s00401-015-1452-x>
27. Larabee CM, Desai S, Agasing A, Georgescu C, Wren JD, Axtell RC, Plafker SM (2016) Loss of Nrf2 exacerbates the visual deficits and optic neuritis elicited by experimental autoimmune encephalomyelitis. *Mol Vis* 22:1503–1513
28. Yu Y, Wu DM, Li J, Deng SH, Liu T, Zhang T, He M, Zhao YY et al (2020) Bixin attenuates experimental autoimmune encephalomyelitis by suppressing TXNIP/NLRP3 inflammasome activity and activating NRF2 signaling. *Front Immunol* 11:593368. <https://doi.org/10.3389/fimmu.2020.593368>
29. Draheim T, Liessem A, Scheld M, Wilms F, Weißflog M, Denecke B, Kensler TW, Zendedel A et al (2016) Activation of the astrocytic Nrf2/ARE system ameliorates the formation of demyelinating lesions in a multiple sclerosis animal model. *Glia* 64(12):2219–2230. <https://doi.org/10.1002/glia.23058>
30. Bresciani G, Manai F, Davinelli S, Tucci P, Saso L, Amadio M (2023) Novel potential pharmacological applications of dimethyl fumarate—an overview and update. *Front Pharmacol* 14:1264842. <https://doi.org/10.3389/fphar.2023.1264842>
31. Michaličková D, Hrnčíř T, Canová NK, Slanař O (2020) Targeting Keap1/Nrf2/ARE signaling pathway in multiple sclerosis. *Eur J Pharmacol* 873:172973. <https://doi.org/10.1016/j.ejphar.2020.172973>
32. Gopal S, Mikulskis A, Gold R, Fox RJ, Dawson KT, Amaravadi L (2017) Evidence of activation of the Nrf2 pathway in multiple sclerosis patients treated with delayed-release dimethyl fumarate in the Phase 3 DEFINE and CONFIRM studies. *Multiple Sclerosis (Houndmills, Basingstoke, England)* 23(14):1875–1883. <https://doi.org/10.1177/1352458517690617>
33. Hammer A, Waschbisch A, Kuhbandner K, Bayas A, Lee DH, Duscha A, Haghikia A, Gold R et al (2018) The NRF2 pathway as potential biomarker for dimethyl fumarate treatment in multiple sclerosis. *Ann Clin Transl Neurol* 5(6):668–676. <https://doi.org/10.1002/acn3.553>
34. Titcomb TJ, Giesser BS, Plafker SM, Katz Sand IB, Wahls TL (2023) Editorial: diet and multiple sclerosis. *Front Neurol* 14:1347478. <https://doi.org/10.3389/fneur.2023.1347478>
35. Pall ML, Levine S (2015) Nrf2, a master regulator of detoxification and also antioxidant, anti-inflammatory and other cytoprotective mechanisms, is raised by health promoting factors. Sheng Li Xue Bao: [Acta Physiologica Sinica] 67(1):1–18
36. Katz Sand I (2018) The role of diet in multiple sclerosis: mechanistic connections and current evidence. *Curr Nutr Rep* 7(3):150–160. <https://doi.org/10.1007/s13668-018-0236-z>
37. Daley SF, Challa HJ, Uppaluri KR (2025) Paleolithic diet. In: StatPearls. StatPearls Publishing Copyright © 2025, StatPearls Publishing LLC., Treasure Island (FL),
38. Lu Y, Yang YY, Zhou MW, Liu N, Xing HY, Liu XX, Li F (2018) Ketogenic diet attenuates oxidative stress and inflammation after spinal cord injury by activating Nrf2 and suppressing the NF- $\kappa$ B signaling pathways. *Neurosci Lett* 683:13–18. <https://doi.org/10.1016/j.neulet.2018.06.016>
39. Milder JB, Liang LP, Patel M (2010) Acute oxidative stress and systemic Nrf2 activation by the ketogenic diet. *Neurobiol Dis* 40(1):238–244. <https://doi.org/10.1016/j.nbd.2010.05.030>
40. Dai S, Wei J, Zhang H, Luo P, Yang Y, Jiang X, Fei Z, Liang W et al (2022) Intermittent fasting reduces neuroinflammation in intracerebral hemorrhage through the Sirt3/Nrf2/HO-1 pathway. *J Neuroinflammation* 19(1):122. <https://doi.org/10.1186/s12974-022-02474-2>
41. Colombo E, Righini M, Balendra V, Rustamli K, Garrone O, Ratti M, Ghidini M (2025) Impact of ketogenic and fast-mimicking diet in gastrointestinal cancer treatment. *Front Oncol* 15:1677509. <https://doi.org/10.3389/fonc.2025.1677509>
42. Espinoza SE, Park S, Connolly G, Qi W, Zhang N, Semwal M, Li Y, Lauzon M et al (2025) Effect of fasting-mimicking diet on markers of autophagy and metabolic health in human subjects. *Geroscience*. <https://doi.org/10.1007/s11357-025-02035-4>
43. Andrews E, Cheng K, Vanderpool C (2022) Nutritional deficiencies in vegetarian, gluten-free, and ketogenic diets. *Pediatr Rev* 43(2):61–70. <https://doi.org/10.1542/pir.2020-004275>
44. Titcomb TJ, Bisht B, Moore DD 3rd, Chhonker YS, Murry DJ, Snetselaar LG, Wahls TL (2020) Eating pattern and nutritional risks among people with multiple sclerosis following a modified Paleolithic diet. *Nutrients*. <https://doi.org/10.3390/nu12061844>
45. Gallop MR, Vieira RFL, Mower PD, Matsuzaki ET, Liou W, Smart FE, Roberts S, Evason KJ et al (2025) A long-term ketogenic diet causes hyperlipidemia, liver dysfunction, and

- glucose intolerance from impaired insulin secretion in mice. *Sci Adv* 11(38):eadx2752. <https://doi.org/10.1126/sciadv.adx2752>
46. Fonseca-Kelly Z, Nassrallah M, Uribe J, Khan RS, Dine K, Dutt M, Shindler KS (2012) Resveratrol neuroprotection in a chronic mouse model of multiple sclerosis. *Front Neurol* 3:84. <https://doi.org/10.3389/fneur.2012.00084>
  47. Chi F, Cheng C, Zhang M, Su B, Hou Y, Bai G (2024) Resveratrol targeting NRF2 disrupts the binding between KEAP1 and NRF2-DLG motif to ameliorate oxidative stress damage in mice pulmonary infection. *J Ethnopharmacol* 332:118353. <https://doi.org/10.1016/j.jep.2024.118353>
  48. Kong D, Yan Y, He XY, Yang H, Liang B, Wang J, He Y, Ding Y et al (2019) Effects of resveratrol on the mechanisms of antioxidants and estrogen in Alzheimer's disease. *Biomed Res Int* 2019:8983752. <https://doi.org/10.1155/2019/8983752>
  49. Ma Y, Zhang J, Zhou Q, Tang P, Wang X, Zhu H, Huang Z, Su H et al (2026) The activation of SIRT1-Nrf2 axis exerts beneficial effects against rotenone-induced cognitive deficits in mice through inhibition of neuroinflammation and ferroptosis. *Ecotoxicol Environ Saf* 309:119697. <https://doi.org/10.1016/j.ecoenv.2026.119697>
  50. Xu JJ, Cui J, Lin Q, Chen XY, Zhang J, Gao EH, Wei B, Zhao W (2021) Protection of the enhanced Nrf2 deacetylation and its downstream transcriptional activity by SIRT1 in myocardial ischemia/reperfusion injury. *Int J Cardiol* 342:82–93. <https://doi.org/10.1016/j.ijcard.2021.08.007>
  51. Hui Y, Chengyong T, Cheng L, Haixia H, Yuanda Z, Weihua Y (2018) Resveratrol attenuates the cytotoxicity induced by Amyloid- $\beta$ (1–42) in PC12 cells by upregulating heme oxygenase-1 via the PI3K/Akt/Nrf2 pathway. *Neurochem Res* 43(2):297–305. <https://doi.org/10.1007/s11064-017-2421-7>
  52. Chen CY, Jang JH, Li MH, Surh YJ (2005) Resveratrol upregulates heme oxygenase-1 expression via activation of NF-E2-related factor 2 in PC12 cells. *Biochem Biophys Res Commun* 331(4):993–1000. <https://doi.org/10.1016/j.bbrc.2005.03.237>
  53. Dinkova-Kostova AT, Talalay P (1999) Relation of structure of curcumin analogs to their potencies as inducers of Phase 2 detoxification enzymes. *Carcinogenesis* 20(5):911–914. <https://doi.org/10.1093/carcin/20.5.911>
  54. Scapagnini G, Colombrita C, Amadio M, D'Agata V, Arcelli E, Sapienza M, Quattrone A, Calabrese V (2006) Curcumin activates defensive genes and protects neurons against oxidative stress. *Antioxid Redox Signal* 8(3–4):395–403. <https://doi.org/10.1089/ars.2006.8.395>
  55. Balogun E, Hoque M, Gong P, Killeen E, Green CJ, Foresti R, Alam J, Motterlini R (2003) Curcumin activates the haem oxygenase-1 gene via regulation of Nrf2 and the antioxidant-responsive element. *Biochem J* 371(Pt 3):887–895. <https://doi.org/10.1042/bj20021619>
  56. McNally SJ, Harrison EM, Ross JA, Garden OJ, Wigmore SJ (2007) Curcumin induces heme oxygenase 1 through generation of reactive oxygen species, p38 activation and phosphatase inhibition. *Int J Mol Med* 19(1):165–172
  57. Thangapazham RL, Sharma A, Maheshwari RK (2006) Multiple molecular targets in cancer chemoprevention by curcumin. *AAPS J* 8(3):E443–449. <https://doi.org/10.1208/aapsj080352>
  58. Xu J, Zhou L, Weng Q, Xiao L, Li Q (2019) Curcumin analogues attenuate  $\text{A}\beta$ (25–35)-induced oxidative stress in PC12 cells via Keap1/Nrf2/HO-1 signaling pathways. *Chem Biol Interact* 305:171–179. <https://doi.org/10.1016/j.cbi.2019.01.010>
  59. Petracca M, Quarantelli M, Moccia M, Vacca G, Satelliti B, D'Ambrosio G, Carotenuto A, Ragucci M et al (2021) Prospective study to evaluate efficacy, safety and tolerability of dietary supplement of Curcumin (BCM95) in subjects with active relapsing Multiple Sclerosis treated with subcutaneous Interferon beta 1a 44 mcg TIW (CONTAIN): A randomized, controlled trial. *Mult Scler Relat Disord* 56:103274. <https://doi.org/10.1016/j.msard.2021.103274>
  60. Niu L, Luo Y, Xie W, Wang C, Liu Z (2026) Dietary (-)-Epigallocatechin gallate (EGCG): state-of-the-art advances in bioactivities, bioavailability enhancement strategies, and applications in nutrition and health. *Nutrients*. <https://doi.org/10.3390/nu18020317>
  61. Romeo L, Intrieri M, D'Agata V, Mangano NG, Oriani G, Ontario ML, Scapagnini G (2009) The major green tea polyphenol, (-)-epigallocatechin-3-gallate, induces heme oxygenase in rat neurons and acts as an effective neuroprotective agent against oxidative stress. *J Am Coll Nutr* 28(Suppl):492s–499s. <https://doi.org/10.1080/07315724.2009.10718116>
  62. He F, Zhang Y, Chen S, Ye B, Chen J, Li C (2018) Effect of EGCG on oxidative stress and Nrf2/HO-1 pathway in neurons exposed to oxygen-glucose deprivation/reperfusion. *Zhong Nan Da Xue Xue Bao Yi Xue Ban* 43(10):1041–1047. <https://doi.org/10.11817/j.issn.1672-7347.2018.10.001>
  63. Kim SR, Seong KJ, Kim WJ, Jung JY (2022) Epigallocatechin gallate protects against hypoxia-induced inflammation in microglia via NF- $\kappa$ B suppression and Nrf-2/HO-1 activation. *Int J Mol Sci*. <https://doi.org/10.3390/ijms23074004>
  64. Na HK, Kim EH, Jung JH, Lee HH, Hyun JW, Surh YJ (2008) (-)-Epigallocatechin gallate induces Nrf2-mediated antioxidant enzyme expression via activation of PI3K and ERK in human mammary epithelial cells. *Arch Biochem Biophys* 476(2):171–177. <https://doi.org/10.1016/j.abb.2008.04.003>
  65. Mi Y, Zhang W, Tian H, Li R, Huang S, Li X, Qi G, Liu X (2018) EGCG evokes Nrf2 nuclear translocation and dampens PTP1B expression to ameliorate metabolic misalignment under insulin resistance condition. *Food Funct* 9(3):1510–1523. <https://doi.org/10.1039/c7fo01554b>
  66. Liu N, Kan QC, Zhang XJ, Xv YM, Zhang S, Zhang GX, Zhu L (2014) Upregulation of immunomodulatory molecules by matrine treatment in experimental autoimmune encephalomyelitis. *Exp Mol Pathol* 97(3):470–476. <https://doi.org/10.1016/j.yexmp.2014.10.004>
  67. Khan A, Shal B, Khan AU, Bibi T, Islam SU, Baig MW, Haq IU, Ali H et al (2021) Withametelin, a novel phytosterol, alleviates neurological symptoms in EAE mouse model of multiple sclerosis via modulation of Nrf2/HO-1 and TLR4/NF- $\kappa$ B signaling. *Neurochem Int* 151:105211. <https://doi.org/10.1016/j.neuint.2021.105211>
  68. Nasrmezhad R, Halalkhor S, Sadeghi F, Pourabdolhossein F (2021) Piperine improves experimental autoimmune encephalomyelitis (EAE) in Lewis rats through its neuroprotective, anti-inflammatory, and antioxidant effects. *Mol Neurobiol* 58(11):5473–5493. <https://doi.org/10.1007/s12035-021-02497-5>
  69. Li B, Cui W, Liu J, Li R, Liu Q, Xie XH, Ge XL, Zhang J et al (2013) Sulforaphane ameliorates the development of experimental autoimmune encephalomyelitis by antagonizing oxidative stress and Th17-related inflammation in mice. *Exp Neurol* 250:239–249. <https://doi.org/10.1016/j.expneurol.2013.10.002>
  70. Devi A, Kabra A, Saeedan AS, Ansari MN (2025) Exploring myricetin: a comprehensive review of its pharmacological potential, formulation strategies, and clinical outlook. *Nahrungsmittelforschung Arch Pharmacol*. <https://doi.org/10.1007/s00210-025-04648-0>
  71. Tancreda G, Ravera S, Panfoli I (2024) Exploring the therapeutic potential: bioactive molecules and dietary interventions in multiple sclerosis management. *Curr Issues Mol Biol* 46(6):5595–5613. <https://doi.org/10.3390/cimb46060335>
  72. Song Y, Wang M, Zhao S, Tian Y, Zhang C (2022) Matrine promotes mitochondrial biosynthesis and reduces oxidative stress in

- experimental optic neuritis. *Front Pharmacol* 13:936632. <https://doi.org/10.3389/fphar.2022.936632>
73. Zhang Q, Li Z, Wu S, Li X, Sang Y, Li J, Niu Y, Ding H (2016) Myricetin alleviates cuprizone-induced behavioral dysfunction and demyelination in mice by Nrf2 pathway. *Food Funct* 7(10):4332–4342. <https://doi.org/10.1039/c6fo00825a>
  74. Khor TO, Fuentes F, Shu L, Paredes-Gonzalez X, Yang AY, Liu Y, Smiraglia DJ, Yegnasubramanian S et al (2014) Epigenetic DNA methylation of antioxidative stress regulator NRF2 in human prostate cancer. *Cancer Prev Res Phila* 7(12):1186–1197. <https://doi.org/10.1158/1940-6207.Capr-14-0127>
  75. Zhao F, Zhang J, Chang N (2018) Epigenetic modification of Nrf2 by sulforaphane increases the antioxidative and anti-inflammatory capacity in a cellular model of Alzheimer's disease. *Eur J Pharmacol* 824:1–10. <https://doi.org/10.1016/j.ejphar.2018.01.046>
  76. Ashrafizadeh M, Ahmadi Z, Mohammadinejad R, Farkhondeh T, Samarghandian S (2020) Curcumin activates the Nrf2 pathway and induces cellular protection against oxidative injury. *Curr Mol Med* 20(2):116–133. <https://doi.org/10.2174/1566524019666191016150757>
  77. Roshanbakhsh H, Elahdadi Salmani M, Dehghan S, Nazari A, Javan M, Pourabdolhossein F (2020) Piperine ameliorated memory impairment and myelin damage in lysoclethrin induced hippocampal demyelination. *Life Sci* 253:117671. <https://doi.org/10.1016/j.lfs.2020.117671>
  78. Feng F, Li X, Wang W, Dou M, Li S, Jin X, Chu Y, Zhu L (2024) Matrine protects against experimental autoimmune encephalomyelitis through modulating microglial ferroptosis. *Biochem Biophys Res Commun* 735:150651. <https://doi.org/10.1016/j.bbrc.2024.150651>
  79. Bhattacharjee S, Dashwood RH (2020) Epigenetic regulation of NRF2/KEAP1 by phytochemicals. *Antioxidants*. <https://doi.org/10.3390/antiox9090865>
  80. Yang X, Liu Y, Cao J, Wu C, Tang L, Bian W, Chen Y, Yu L et al (2025) Targeting epigenetic and post-translational modifications of NRF2: key regulatory factors in disease treatment. *Cell Death Discov* 11(1):189. <https://doi.org/10.1038/s41420-025-02491-z>
  81. Moraca F, Arciuolo V, Marzano S, Napolitano F, Castellano G, D'Aria F, Di Porzio A, Landolfi L et al (2025) Repurposing FDA-approved drugs to target G-quadruplexes in breast cancer. *Eur J Med Chem* 285:117245. <https://doi.org/10.1016/j.ejmech.2025.117245>
  82. Napolitano F, Di Somma S, Castellano G, Amato J, Pagano B, Randazzo A, Portella G, Malfitano AM (2022) Combination of dl922-947 oncolytic adenovirus and G-quadruplex binders uncovers improved antitumor activity in breast cancer. *Cells*. <https://doi.org/10.3390/cells11162482>
  83. Di Somma S, Amato J, Iaccarino N, Pagano B, Randazzo A, Portella G, Malfitano AM (2019) G-quadruplex binders induce immunogenic cell death markers in aggressive breast cancer cells. *Cancers (Basel)* 11(11). <https://doi.org/10.3390/cancers1111797>
  84. Milošević M, Arsić A, Cvetković Z, Vučić V (2021) Memorable food: fighting age-related neurodegeneration by precision nutrition. *Front Nutr* 8:688086. <https://doi.org/10.3389/fnut.2021.688086>
  85. Chen M, Yang LL, Hu ZW, Qin C, Zhou LQ, Duan YL, Bosco DB, Wu LJ et al (2020) Deficiency of microglial Hv1 channel is associated with activation of autophagic pathway and ROS production in LPC-induced demyelination mouse model. *J Neuroinflammation* 17(1):333. <https://doi.org/10.1186/s12974-020-02020-y>
  86. Moruno-Manchon JF, Lejault P, Wang Y, McCauley B, Honarpisheh P, Morales Scheihing DA, Singh S, Dang W, et al. (2020) Small-molecule G-quadruplex stabilizers reveal a novel pathway of autophagy regulation in neurons. *Elife* 9. <https://doi.org/10.7554/eLife.52283>
  87. Rubio K, Hernández-Cruz EY, Rogel-Ayala DG, Sarvari P, Isidoro C, Barreto G, Pedraza-Chaverri J (2023) Nutriepigenomics in environmental-associated oxidative stress. *Antioxidants*. <https://doi.org/10.3390/antiox12030771>
  88. Waller ZA, Howell LA, Macdonald CJ, O'Connell MA, Searcey M (2014) Identification and characterisation of a G-quadruplex forming sequence in the promoter region of nuclear factor (erythroid-derived 2)-like 2 (Nrf2). *Biochem Biophys Res Commun* 447(1):128–132. <https://doi.org/10.1016/j.bbrc.2014.03.117>
  89. Lee SC, Zhang J, Strom J, Yang D, Dinh TN, Kappeler K, Chen QM (2017) G-quadruplex in the NRF2 mRNA 5' untranslated region regulates de novo NRF2 protein translation under oxidative stress. *Mol Cell Biol*. <https://doi.org/10.1128/mcb.00122-16>
  90. Malfitano AM, Castellano G, Croce A, Napolitano F, Portella G, Beguinot F, Formisano P, Fiory F (2025) Nutritional bioactive compounds with beneficial effects for multiple sclerosis: potential implication of G-Quadruplexes? *Mutat Res Rev Mutat Res* 796:108548. <https://doi.org/10.1016/j.mrrev.2025.108548>
  91. Tawani A, Amanullah A, Mishra A, Kumar A (2016) Evidences for piperine inhibiting cancer by targeting human G-quadruplex DNA sequences. *Sci Rep* 6:39239. <https://doi.org/10.1038/srep39239>
  92. Ye H, Zhang H, Xiang J, Shen G, Yang F, Wang F, Wang J, Tang Y (2024) Advances and prospects of natural dietary polyphenols as G-quadruplex stabilizers in biomedical applications. *Int J Biol Macromol* 254(2):127825. <https://doi.org/10.1016/j.ijbiomac.2023.127825>
  93. Storoni M, Plant GT (2015) The therapeutic potential of the ketogenic diet in treating progressive multiple sclerosis. *Mult Scler Int* 2015:681289. <https://doi.org/10.1155/2015/681289>
  94. Benlloch M, López-Rodríguez MM, Cuerda-Ballester M, Drehmer E, Carrera S, Ceron JJ, Tvarijonaviciute A, Chirivella J, et al. (2019) Satiating effect of a ketogenic diet and its impact on muscle improvement and oxidation state in multiple sclerosis patients. *Nutrients* 11 (5). <https://doi.org/10.3390/nu11051156>
  95. Brenton JN, Banwell B, Bergqvist AGC, Lehner-Gulotta D, Gampper L, Leytham E, Coleman R, Goldman MD (2019) Pilot study of a ketogenic diet in relapsing-remitting MS. *Neurol Neuroimmunol Neuroinflamm* 6(4):e565. <https://doi.org/10.1212/nxi.0000000000000565>
  96. Ordoñez-Rodríguez A, Roman P, Rueda-Ruzafa L, Campos-Rios A, Cardona D (2023) Changes in gut microbiota and multiple sclerosis: a systematic review. *Int J Environ Res Public Health* 20(5). <https://doi.org/10.3390/ijerph20054624>
  97. Swidsinski A, Dörffel Y, Loening-Baucke V, Gille C, Göktas Ö, Reißhauer A, Neuhaus J, Weylandt KH et al (2017) Reduced mass and diversity of the colonic microbiome in patients with multiple sclerosis and their improvement with ketogenic diet. *Front Microbiol* 8:1141. <https://doi.org/10.3389/fmicb.2017.01141>
  98. Willcox DC, Scapagnini G, Willcox BJ (2014) Healthy aging diets other than the Mediterranean: a focus on the Okinawan diet. *Mech Ageing Dev* 136:148–162. <https://doi.org/10.1016/j.mad.2014.01.002>
  99. Hao M, Tan X, Liu K, Xin N (2025) Nanoencapsulation of nutraceuticals: enhancing stability and bioavailability in functional foods. *Front Nutr* 12:1746176. <https://doi.org/10.3389/fnut.2025.1746176>
  100. Aliyu M, Saboor-Yaraghi AA, Sahraian MA, Noorbakhsh F (2025) The experimental autoimmune encephalomyelitis (EAE) model: a gateway to successful translation of multiple sclerosis

- therapies. *Iran J Allergy Asthma Immunol* 24(5):563–580. <https://doi.org/10.18502/ijaai.v24i5.19740>
101. Swank RL (1953) Treatment of multiple sclerosis with low-fat diet. *AMA Arch Neurol Psychiatry* 69(1):91–103. <https://doi.org/10.1001/archneurpsyc.1953.02320250097011>
102. Bisht B, Darling WG, Grossmann RE, Shivapour ET, Lutgen-dorf SK, Snetselaar LG, Hall MJ, Zimmerman MB et al (2014) A multimodal intervention for patients with secondary progressive multiple sclerosis: feasibility and effect on fatigue. *J Altern Complement Med* 20(5):347–355. <https://doi.org/10.1089/acm.2013.0188>
103. Irish AK, Erickson CM, Wahls TL, Snetselaar LG, Darling WG (2017) Randomized control trial evaluation of a modified Paleolithic dietary intervention in the treatment of relapsing-remitting multiple sclerosis: a pilot study. *Degener Neurol Neuromuscul Dis* 7:1–18. <https://doi.org/10.2147/dmnd.S116949>
104. Dean Z, Penesova A, Radikova Z, Dean Z, Kollar B (2022) Pilot study of longterm low fat diet in relapsing-remitting multiple sclerosis. *Neuro Endocrinol Lett* 43(3):135–139
105. Swank RL, Goodwin J (2003) Review of MS patient survival on a Swank low saturated fat diet. *Nutrition* 19(2):161–162. [https://doi.org/10.1016/s0899-9007\(02\)00851-1](https://doi.org/10.1016/s0899-9007(02)00851-1)
106. Haß U, Herpich C, Norman K (2019) Anti-inflammatory diets and fatigue. *Nutrients*. <https://doi.org/10.3390/nu11102315>
107. Moravejolahkami AR, Chitsaz A, Hassanzadeh A, Paknahad Z (2023) Effects of anti-inflammatory-antioxidant-rich diet and co-supplemented synbiotics intervention in patients with progressive forms of multiple sclerosis: a single-center, single-blind randomized clinical trial. *Nutr Neurosci* 26(11):1078–1089. <https://doi.org/10.1080/1028415x.2022.2128010>
108. Alfredsson L, Olsson T, Hedström AK (2023) Inverse association between Mediterranean diet and risk of multiple sclerosis. *Multiple Sclerosis (Houndmills, Basingstoke, England)* 29(9):1118–1125. <https://doi.org/10.1177/13524585231181841>
109. Katz Sand I, Benn EKT, Fabian M, Fitzgerald KC, Digga E, Deshpande R, Miller A, Gallo S et al (2019) Randomized-controlled trial of a modified Mediterranean dietary program for multiple sclerosis: a pilot study. *Multiple Sclerosis Relat Disord* 36:101403. <https://doi.org/10.1016/j.msard.2019.101403>
110. Razeghi-Jahromi S, Doosti R, Ghorbani Z, Saeedi R, Abolhasani M, Akbari N, Cheraghi-Serkani F, Moghadasi AN et al (2020) A randomized controlled trial investigating the effects of a mediterranean-like diet in patients with multiple sclerosis-associated cognitive impairments and fatigue. *Curr J Neurol* 19(3):112–121. <https://doi.org/10.18502/cjn.v19i3.5424>
111. Bahr LS, Bellmann-Strobl J, Koppold DA, Rust R, Schmitz-Hüb-sch T, Olszewska M, Stadlbauer J, Bock M et al (2025) Fasting, ketogenic, and anti-inflammatory diets in multiple sclerosis: a randomized controlled trial with 18-month follow-up. *BMC Nutr* 11(1):167. <https://doi.org/10.1186/s40795-025-01156-5>
112. Keramatzadeh S, Hosseini SA, Majdinasab N, Cheraghian B, Zilaei M (2025) Effects of resveratrol supplementation on inflammatory markers, fatigue scale, fasting blood sugar and lipid profile in relapsing-remitting multiple sclerosis patients: a double-blind, randomized placebo-controlled trial. *Nutr Neurosci* 28(7):854–862. <https://doi.org/10.1080/1028415x.2024.2425649>

**Publisher's Note** Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.