



Frequency and clinical correlates of anti-nerve antibodies in a large population of CIDP patients included in the Italian database

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Abstract

Objective To investigate the frequency and clinical correlates of anti-nerve autoantibodies in an unselected series of Italian patients with chronic inflammatory demyelinating polyradiculoneuropathy (CIDP)

Methods Sera from 276 CIDP patients fulfilling the EFNS/PNS criteria and included in the Italian CIDP database were examined for the presence of anti-nerve autoantibodies. Results were correlated with the clinical data collected in the database.

Results Anti-neurofascin155 (NF155) antibodies were found in 9/258 (3.5%) patients, anti-contactin1 (CNTN1) antibodies in 4/258 (1.6%) patients, and anti-contactin-associated protein1 (Caspr1) in 1/197 (0.5%) patients, while none had reactivity to gliomedin or neurofascin 186. Predominance of IgG4 isotype was present in 7 of the 9 examined patients. Anti-NF155 patients more frequently had ataxia, tremor, and higher CSF protein levels than antibody-negative patients. Anti-CNTN1 patients more frequently had a GBS-like onset, pain, and ataxia and had more severe motor impairment at enrollment than antibody-negative patients. They more frequently received plasmapheresis, possibly reflecting a less satisfactory response to IVIg or steroids. IgM antibodies against one or more gangliosides were found in 6.5% of the patients (17/260) and were more frequently directed against GM1 (3.9%). They were frequently associated with a progressive course, with a multifocal sensorimotor phenotype and less frequent cranial nerve involvement and ataxia.

Conclusions Anti-paranodal and anti-ganglioside antibodies are infrequent in patients with CIDP but are associated with some typical clinical association supporting the hypothesis that CIDP might be a pathogenically heterogeneous syndrome possibly explaining the different clinical presentations.

Keywords Chronic inflammatory demyelinating polyradiculoneuropathy · CIDP · Peripheral neuropathy · Anti-nerve antibodies · paranodopathy · anti-ganglioside antibodies

Introduction

Chronic inflammatory demyelinating polyneuropathy (CIDP) is the most frequent chronic immune-mediated neuropathy with a prevalence ranging from 0.67 to 8.9 cases per 100 000 [1]. Despite the elusiveness of its exact pathogenic

mechanism [2], the immunological involvement in CIDP is supported by its frequent improvement after immune therapies [3]. The disease course can be either relapsing-remitting or progressive and is typically characterized by a symmetric sensorimotor involvement although several variants have been described broadening the spectrum of this disorder [4].

The identification of disease-associated antibodies in other neuropathies has already entered clinical practice reshaping their clinical management and treatment strategies [5]. A number of recent studies have identified a few reactivities in CIDP patients against cell adhesion molecules at the paranodal (neurofascin-155 [6–14], contactin-1 [12, 13, 15–17], contactin-associated protein 1 [13, 18]), or nodal

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domain (neurofascin 186 [13, 14], gliomedin) or against membrane gangliosides [19–21]. Despite the variable prevalence of these reactivities in different studies, there is some evidence that some of these reactivities are associated with some typical clinical features and response to therapy.

In the present study, we assessed the prevalence of anti-nerve antibodies in a large and consecutive series of Italian CIDP patients to provide further information regarding the prevalence and the clinical correlates of these antibodies.

Material and methods

Patients and samples

This was a retrospective multicenter cohort study on large series of CIDP patients collected in the Italia CIDP database (CINECA, Bologna, Italy). The details of this study have been previously reported [22]. Clinical and diagnostic data were consecutively collected in each center from all currently followed patients with a diagnosis of CIDP and independently from therapy response. The diagnostic accuracy was centrally verified according to the EFNS/PNS diagnostic criteria [23]. Data monitoring included diagnosis revision, suspect double entries, missing data, and plausibility

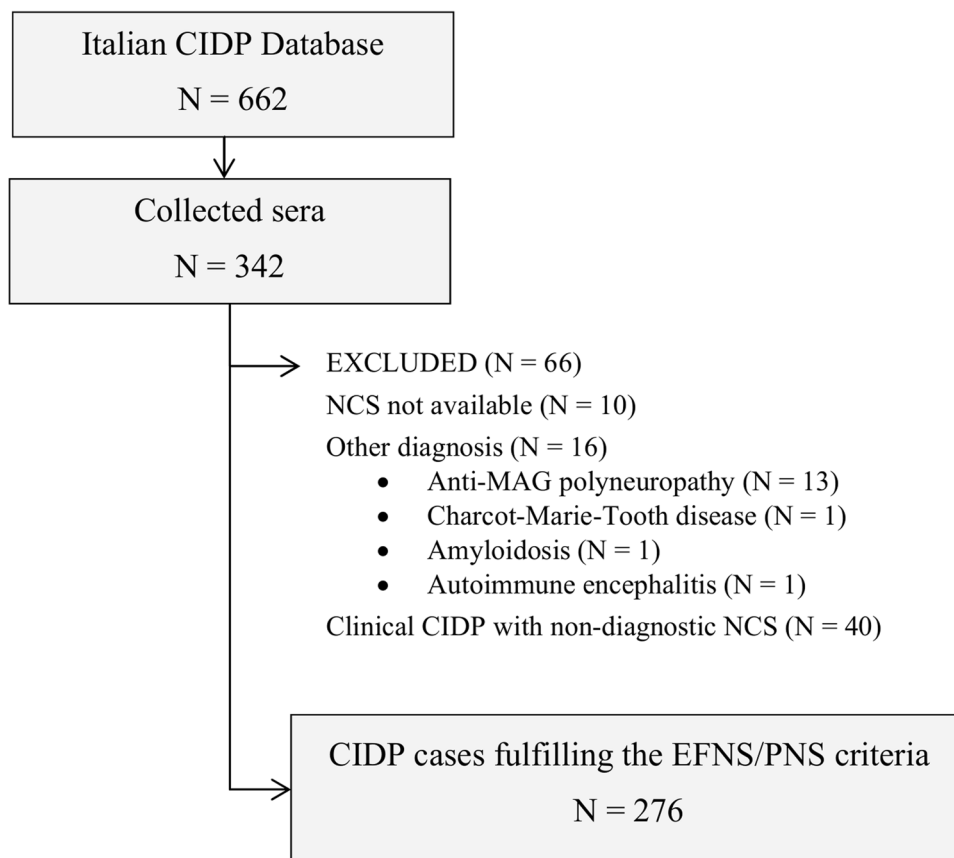
checks. We excluded patients with an alternative diagnosis, IgM monoclonal gammopathy, and increased titers of anti-myelin-associated glycoprotein (MAG) IgM antibodies (over 7000 Units by the Buhlman method in our laboratory [24]), increased levels of circulating VEGF (> 1500 pg/mL) [25], unavailable nerve conduction studies (NCS), or data not fulfilling the EFNS/PNS diagnostic criteria [23]. The study was approved by the Ethical Committee of each participating center. All patients gave written informed consent.

Among the 662 patients included in the database, sera were only provided by 15 participating centers who provided the sera from all their included patients for a total of 342 patients. After revision of the diagnosis, we excluded 16 patients with an alternative diagnosis, 10 patients with unavailable NCS, and 40 patients not fulfilling the EFNS/PNS electrodiagnostic criteria leading to a final study population of 276 patients (Fig. 1).

Serological analysis

Anti-node/paranode antibody testing Antibodies were measured by ELISA according to previously reported procedure [7, 15]. Briefly, 96-well Nunc Polysorb ELISA plates were coated overnight at 4 °C with 1 µg/mL of human recombinant NF155 protein (OriGene RC228652) or human

Fig. 1 Flow-chart of the case population



recombinant CNTN1 protein (OriGene RC214706). Wells were saturated with 5% non-fat milk in 0.1% PBS-Tween 20 solution for 1 h at room temperature (RT) and then incubated in duplicate with sera diluted 1:100 in saturating solution for 1 h at RT. Horseradish peroxidase–labeled polyclonal rabbit anti-human IgG/HRP was added at the dilution of 1:10,000 in saturating solution for 1 h at RT. For antibodies to NF186 and Caspr1, plates were coated with 1 µg/mL human recombinant NF186 protein (TP 329070 OriGene) or 5 µg/mL human recombinant Caspr1 protein (2418-CR R&D) at the same serum dilution while the polyclonal rabbit anti-human IgG/HRP was diluted 1:1000 in saturating. IgG subclasses 1–4 were determined using the appropriate horseradish peroxidase–conjugated mouse-anti-human IgG with (Life Technologies) diluted 1:500 in saturating solution. Reactivity was detected with TMB solution (BioLegend), and the reaction was stopped with 0.1M sulfuric acid. Optical density (OD) was measured at a wavelength of 450 nm by a DSX plate reader (manufactured by Technogenetics). Only patients with an optical density > 0.3 had their positivity confirmed in all by immunocytochemistry on transfected human embryonic kidney (HEK) 293 cells at the Neuromuscular Laboratory of the Neurology Department, Hospital de la Santa Creu i Sant Pau, Barcelona, Spain (Dr. Luis Querol) [7, 15].

Anti-ganglioside IgM antibodies All collected sera were tested by ELISA for the presence of anti-ganglioside IgM antibodies (anti-GAAb) by individual assay against GM1, GM2, GD1a, GD1b, and GQ1b using previously reported procedures [26] with an upper normal limit for serum antibody reactivity of 1/640.

Clinical features

All patients had a detailed clinical history including duration of weakness, sensory symptoms, ataxia, pain, and autonomic dysfunction. The course of the disease was defined by the treating neurologist as progressive or relapsing and an eventual GBS-like onset was reported [22]. Response to previously performed therapy was reported by the treating neurologist and defined as an improvement of at least 2 points at the Medical Research Council (MRC) score (range 0, worst–60, normal) or at least 1 point on the INCAT scale (range 0, normal–10, worst). The clinical evaluation at entry also included the INCAT sensory sum score (ISS), range 1 (normal)–20 (worst) [27]. Results of cerebrospinal fluid (CSF) examination performed during the course of the disease were reported. The upper reference limit for CSF proteins was considered 50 mg/dL for patients aged ≤ 50 years and 60 mg/dL for those aged > 50 years [28]. Motor nerve conduction studies were planned to be performed bilaterally in the median, ulnar, common peroneal, and tibial nerves

and included distal and proximal compound muscle action potential (CMAP) amplitude (onset to peak) and duration, motor conduction velocities (MCV), distal and proximal motor latencies, and in most patients F-wave latency. The results were centrally reviewed and classified according to the EFNS/PNS criteria [23].

Statistical analysis

Categorical variables were described using frequency and percentage and analyzed with the chi-square or Fisher exact tests. Continuous variables were described using mean and standard deviation, assessed for normality with the Shapiro-Wilk test and analyzed with the *t*-test (for normally distributed variables) or Wilcoxon-Mann-Whitney test (for non-parametrically distributed variables). Significance was set at an α -level of 0.05; no multiple testing correction was applied. Analyses were performed with IBM SPSS Statistics for Windows, Version 22.0 (Armonk, NY: IBM Corp, USA).

Results

Anti-node/paranode IgG antibodies

Anti-NF155 and CNTN1 IgG antibodies were measured in 258 patients, and result increased in 9 patients for anti-NF155 IgG (3.5%) and four for anti-CNTN1 IgG (1.6%). Sera from 197 patients were tested for anti-Caspr1, anti-NF186, and anti-gliomedin IgG antibodies with reactivity observed in one patient for Caspr1 (0.5%) and none for NF186 or gliomedin. Antibody subtype analysis was performed in five of the nine patients with anti-NF155 antibodies and resulted positive for IgG4 in all but one patient in whom isotype characterization was not conclusive. Subtype analysis in the four anti-CNTN1-positive sera resulted positive for IgG4 in all patients with one also having IgG3 antibodies. Follow-up serological analysis in the two patients with anti-NF155 examined after therapy revealed a decrease of antibody reactivity with an OD reduction from 1.576 to 0.900 in one and from 1.296 to 0.327 in the other in parallel with clinical improvement. In the three patients with anti-CNTN1 examined after therapy, antibody decreased from 0.678, 0.775, and 0.839 of OD to 0.100 or less paralleling clinical improvement.

Comparison of clinical features of patients with and without anti-paranodal antibodies

In Table 1 are compared the clinical, diagnostic, and therapeutic findings in patients with anti-NF155, anti-CNTN1 antibodies, and without any of these antibodies.

Patients with anti-NF155 population had a shorter disease duration at enrollment (40.44 vs. 47.41 years) and had more frequently tremor, ataxia, and cranial nerve involvement,

Table 1 Comparison of characteristics of patients with anti-NF155 IgG or anti-CNTN1 IgG and seronegative CIDP patients

	Anti-NF155 (n = 9)	Anti-CNTN1 (n = 4)	Seronegative (n = 245)	P values
Gender, male, n	5/9 (55.6%)	3/4 (75%)	171/245 (70.1%)	
Age at onset, mean (\pm SD)	40.44 (\pm 23.61)	52.5 (\pm 15.15)	47.41 (\pm 16.84)	
Disease duration at enrollment, mean (\pm SD)	5.02 (\pm 6.30)	4.8 (\pm 5.43)	9.51 (\pm 8.80)	$p^1 = 0.026$
CIDP subtype, n				
Typical CIDP	9/9 (100%)	4/4 (100%)	175/245 (71.7%)	$p^1 = 0.024$
Atypical CIDP	0/9 (0%)	0/4 (0%)	69/245 (28.3%)	
Disease course, n				
Relapsing	5/9 (55.6%)	2/4 (50%)	115/245 (47.1%)	
Progressive	4/9 (44.4%)	2/4 (50%)	129/245 (52.9%)	
GBS-like onset	1/9 (11.1%)	2/4 (50%)	24/245 (9.8%)	$p^3 = 0.056$
Impairment, mean (\pm SD)				
INCAT	2.25 (\pm 0.71)	6 (\pm 2.94)	2.46 (\pm 2.00)	$p^3 = 0.001$
MRC	55.25 (\pm 3.2)	42.5 (\pm 13.27)	54.83 (\pm 6.66)	$p^3 = 0.0004$
ISS	6.63 (\pm 3.85)	8.25 (\pm 5.91)	4.67 (\pm 3.89)	
Symptoms at onset, n				
Motor	8/9 (88.9%)	3/4 (75%)	152/245 (62.3%)	
Sensory	8/9 (88.9%)	4/4 (100%)	197/245 (80.7%)	
Pain	2/9 (22.2%)	3/4 (75%)	53/245 (21.7%)	$p^3 = 0.037$
Fatigue	1/9 (11.1%)	2/4 (50%)	87/245 (35.7%)	
Cranial symptoms	3/9 (33.3%)	0/4 (0%)	23/245 (9.4%)	$p^2 = 0.053$
Ataxia	4/9 (44.4%)	1/4 (25%)	28/245 (11.5%)	$p^2 = 0.017$
Cramps	1/9 (11.1%)	0/4 (0%)	30/245 (12.3%)	
All symptoms developed, n				
Motor	9/9 (100%)	4/4 (100%)	218/245 (89.3%)	
Sensory	9/9 (100%)	4/4 (100%)	231/245 (94.7%)	
Pain	2/9 (22.2%)	2/4 (50%)	76/245 (31.1%)	
Fatigue	3/9 (33.3%)	2/4 (50%)	125/245 (51.2%)	
Cranial symptoms	3/9 (33.3%)	1/4 (25%)	49/245 (20.1%)	
Ataxia	7/9 (77.8%)	3/4 (75%)	59/245 (24.2%)	$p^2 = 0.001$; $p^3 = 0.049$
Cramps	3/9 (33.3%)	0/4 (0%)	39/245 (16.0%)	
Tremor	6/9 (55.6%)	0/4 (0%)	31/245 (12.7%)	$p^2 = 0.004$
Autonomic symptoms	0/9 (0%)	0/4 (0%)	23/245 (9.4%)	
EMG findings, n				
RCV	6/9 (66.7%)	4/4 (100%)	148/245 (60.7%)	
CB	4/9 (44.4%)	2/4 (50%)	139/245 (57.0%)	
PDL	5/9 (55.6%)	1/4 (25%)	61/245 (25.0%)	
ATD	0/9 (0%)	0/4 (0%)	29/245 (11.9%)	
PFL or AF	0/9 (0%)	0/4 (0%)	24/245 (9.8%)	
CSF analysis				
Protein concentration (mg/dL), mean (\pm SD)	177.75 (\pm 120.91)	186.5 (\pm 114.90)	94.35 (\pm 86.35)	$p^2 = 0.031$; $p^3 = 0.037$
Age-adjusted CSF protein positivity, n	6/8 (75%)	4/4 (100%)	132/182 (72.5%)	
Biopsy, n				
Demyelination	1/1 (100%)	2/2 (100%)	10/14 (71.4%)	
Axonal	0/1 (0%)	0/2 (0%)	4/14 (28.6%)	
Treatment response, n				
IVIg, responder	5/8 (62.5%)	1/4 (25%)	135/188 (71.8%)	$p^3 = 0.0754$
Steroids, responder	4/7 (57.1%)	1/4 (25%)	87/143 (60.8%)	
PE, responder	0/1 (0%)	3/4 (66.7%)	16/22 (72.7%)	
RTX, responder	2/2 (100%)	2/2 (100%)	1/3 (33.3%)	

p^1 Anti-paranode vs. seronegatives

p^2 Anti-NF155 vs. seronegatives

p^3 Anti-CNTN1 vs. seronegatives

Abbreviations: RCV reduced conduction velocity, CB conduction block, PDL prolonged distal latency, ATD abnormal temporal dispersion, PFL prolonged F-wave latency, AF absent F-wave, IVIg intravenous immunoglobulins, PE plasma exchange, RTX rituximab

consisting in dysphagia and dysphonia in two patients and facial hypoesthesia in one. CSF analysis revealed a higher mean protein concentration with a similar frequency of increased CSF proteins. There was no significant difference in the response to IVIg and steroids. Both treated patients with anti-NF155 antibodies improved after therapy with rituximab compared to one of the three without these antibodies.

Patients with anti-CNTN1 antibodies also had a shorter disease duration at enrolment and more frequently had a GBS-like onset. They were more severely affected with a lower mean MRC score, and higher INCAT score. They also had more frequent pain at onset (75% vs. 21.6% $p = 0.037$) and more frequently developed ataxia during the course of the disease. They also had higher mean CSF protein concentration (186.5 vs. 94.35 mg/dL, $p = 0.037$) with a similar frequency of increased CSF proteins (100% vs. 72.5%). They had a less frequent response to IVIg compared to patients without these antibodies and were more likely to receive plasmapheresis (75%) than seronegative patients (9%) with a similarly frequent response. Both treated patients improved after therapy with rituximab.

Reactivity against Caspr1 was only found in a 57-year-old lady with progressive proximal and distal motor involvement of the four limbs followed by paresthesia, ataxia, and subsequent tremor leading to a diagnosis of definite typical CIDP. The patient did not improve after therapy with IVIg alone or in combination to corticosteroids and with subsequent plasma exchange. The patient subsequently stabilized with physiotherapy and is now planning to receive rituximab. Pain was not a prominent symptom for the patient.

Anti-ganglioside IgM-positive CIDP

IgM antibodies to one or more ganglioside were found in 17 of the 260 examined patients (6.5%). Anti-GM1 IgM antibodies were detected in 10/260 patients (3.9%), anti-GM2 in 5/220 (2.3%), anti-GD1a in 3/224 (1.3%), and anti-GD1b in 8/213 (3.8%), while none had anti-GQ1b IgM. There was a concomitant reactivity with GM1 and GD1b in two patients; with GM1 and GM2 in one patient; with GM2 and GD1a in one patient; with GM1, GD1b, and GD1a in one patient; and with all tested antigens in one patient. The median detected antibody reactivity was 1/5120 with a minimum reactivity of 1/1280.

In Table 2, we separately compared patient with any anti-ganglioside antibody (anti-GAAb) or with anti-GM1 antibodies alone or in combination (GM1) with seronegative patients. Patients with anti-GAAb IgM were older at disease onset (54.8 vs. 46.5 years, $p = 0.044$) and more frequently had a progressive course (88.2% vs. 50.4% $p = 0.002$). They were also less frequently diagnosed to have typical CIDP (41.2% vs. 74.2%, $p = 0.009$) and more

frequently had Lewis-Summer syndrome (35.3% vs. 6.9% $p = 0.002$). A similar difference was also observed in patients with anti-GM2 IgM antibodies with typical CIDP in 20% of the patients ($p = 0.019$) and Lewis-Summer syndrome in 60% ($p = 0.005$). Pain was more frequent in patients with anti-GM1 antibodies (60% vs. 25.9%, $p = 0.027$), or anti-GD1a IgM (100% vs. 25.9%, $p = 0.018$) and in the whole anti-GAAb population (52.9% vs. 25.9%, $p = 0.024$), while cranial nerve involvement (0% vs. 22.9%, $p = 0.027$) and ataxia (5.9% vs. 28.5%, $p = 0.047$) were less frequent in patients with anti-GM1 or anti-ganglioside antibodies. Two patients with anti-GM1 antibodies had a pure motor CIDP. In both patients, motor impairment was relatively symmetric without a multineuropathic distribution making it unlikely a diagnosis of multifocal motor neuropathy. One patient had conduction block in motor nerve and responded to IVIg, while the other did not have conduction block and did not improve after IVIg. There was no difference in the response to therapy according to the presence of anti-ganglioside or anti-GM1 antibodies.

Discussion

The reported prevalence of anti-paranodal IgG in previous studies is quite heterogeneous (between 1 and 20.7% for anti-NF155, between 0.7 and 7.5% for anti-CNTN1, and between 0.2 and 2.9% for anti-Caspr1). These discrepancies may reflect differences in case selection, applied diagnostic criteria, duration of disease and treatment status of screened patients [6–18, 29–31]. The prevalence of these antibodies in our series of CIDP patients was lower than previously reported. This may reflect the unselected series of patients examined in our study and the fact the most of them were not treatment-naïve. The latter hypothesis is possibly confirmed by the fact that in all the four patients in whom follow-up serological analysis were available after therapy, there was a marked reduction in antibody titer flanking clinical remission [6, 13, 17], underlining the importance of screening for these autoantibodies before initiating treatment. In our series, IgG subtype analysis revealed a striking predominance of IgG4 with only one case where both IgG3 and IgG4 as also previously reported. The use of ELISA as a screening test and of cell-based assay as a confirmatory test is similar to what has been performed in previous studies [9, 12, 17] and takes advantage both of the higher availability and reproducibility of ELISA and the lower rate of false positives of cell-based assays [11].

Despite the relatively small number of our positive patients, our findings confirm previously described features of the clinical phenotype of anti-NF155-positive patients (younger age at onset, tremor, ataxia), their electrodiagnostic features with frequently increased distal latencies, and high

Table 2 Comparison of characteristics of patients with anti-GAAb IgM and seronegative CIDP patients

	aGAAb (<i>n</i> = 17)	Anti-GM1 (<i>n</i> = 10)	<i>P</i> values
Gender, male, <i>n</i>	14/17 (82.4%)	9/10 (90%)	
Age at onset (years), mean (\pm SD)	54.82 (\pm 13.45)	56.1 (\pm 12.75)	
Disease duration at enrollment (years), mean (\pm SD)	9.85 (\pm 8.01)	11.15 (\pm 8.58)	
CIDP subtype, <i>n</i>			
Typical	7/17 (41.2%)	6/10 (60%)	$p^1 = 0.009$
DADS	1/17 (5.9%)	0/10 (0%)	
Lewis-Summer syndrome	6/17 (35.3%)	1/10 (10%)	$p^1 = 0.002$
Pure motor CIDP	2/17 (11.8%)	2/10 (20%)	
Pure sensory CIDP	1/17 (5.9%)	1/10 (10%)	
Disease course, <i>n</i>			
Relapsing	2/17 (11.8%)	1/10 (10%)	
Progressive	15/17 (88.2%)	9/10 (90%)	$p^1 = 0.003$; $p^2 = 0.020$
Impairment, mean (\pm SD)			
INCAT	2.08 (\pm 1.85)	2.56 (\pm 1.94)	
MRC	55.54 (\pm 5.74)	54.11 (\pm 6.41)	
ISS	4.23 (\pm 3.66)	4 (\pm 4.36)	
All symptoms developed, <i>n</i>			
Motor	16/17 (94.1%)	10/10 (100%)	
Sensory	15/17 (88.23%)	9/10 (90%)	
Pain	9/17 (52.9%)	6/10 (60%)	$p^1 = 0.024$; $p^2 = 0.027$
Fatigue	8/17 (47.1%)	7/10 (70%)	
Cranial symptoms	0/17 (0%)	0/10 (0%)	$p^1 = 0.027$
Ataxia	1/17 (5.9%)	1/10 (10%)	$p^1 = 0.047$
Cramps	4/17 (23.5%)	2/10 (20%)	
Tremor	0/17 (0%)	0/10 (0%)	
Autonomic symptoms	1/17 (5.9%)	1/10 (10%)	
EMG findings, <i>n</i>			
RCV	11/17 (64.7%)	8/10 (80%)	
CB	7/17 (41.2%)	4/10 (40%)	
PDL	4/17 (23.5%)	4/10 (40%)	
ATD	3/17 (17.6%)	2/10 (20%)	
PFL or AF	1/17 (5.9%)	0/10 (0%)	
Treatment response, <i>n</i>			
IVIg, responder	11/13 (84.6%)	6/8 (75%)	
Steroids, responder	2/5 (40%)	1/3 (33.3%)	

p^1 Anti-ganglioside positives vs. seronegatives

p^2 Anti-GM1 positives vs. seronegatives

Abbreviations: *RCV* reduced conduction velocity, *CB* conduction block, *PDL* prolonged distal latency, *ATD* abnormal temporal dispersion, *PFL* prolonged F-wave latency, *AF* absent F-wave, *IVIg* intravenous immunoglobulins

levels of CSF proteins. Similarly, patients with high anti-CNTN1 antibodies had an advanced age at onset, a frequent GBS-like onset, a prominent motor involvement, and high levels of CSF proteins. We also confirmed that these patients had a low rate of response to IVIg even if the data was not statistically different from seronegative patients. This could be also related to administration of IVIg in addition to corticosteroids in several patients recorded as responders or to

the difficulty in differentiating sustained from non-sustained response in our retrospective analysis. Nevertheless, the frequent use of plasmapheresis in the anti-CNTN1 population may reflect a higher prevalence of unsatisfactory responses to first-line therapies since plasmapheresis in our CIDP population was mostly performed in patients failing to respond to steroids or IVIg. All our treated patients improved after therapy with rituximab supporting the role of this therapy

in this group of patients. Only one patient had antibodies to Caspr1 confirming the low prevalence of this reactivity. Pain was not however a prominent feature in this patient who had an otherwise typical severe CIDP poorly responsive to conventional therapies. No IgG reactivity was found against either NF186 or gliomedin coherently with the described rarity of these autoantibodies [6–9, 12–14].

We also found a consistent proportion who had increased titers of anti-ganglioside antibodies mostly directed against GM1 with a high minimum reactivity level, thus increasing our confidence in the identification of this population. The majority of these patients had atypical presentation consistent with Lewis-Summer syndrome, as also previously reported [20], or a pure motor symmetric CIDP. This finding may represent a link between CIDP and multifocal motor neuropathy even if the sensory impairment in those with Lewis-Summer syndrome and the symmetric involvement in those with motor CIDP are not consistent with this diagnosis. This finding supports, however, the opportunity to test for these antibodies in patients with atypical variant of CIDP. A similar lower rate of cranial nerve involvement was also previously reported in CIDP patients with anti-LM1 antibodies [32].

Even if this study reveals the presence of one or more anti-neural antibodies in 12% of the examined patients, it does not provide additional data that might support their possible pathogenetic relevance. Their correlation with some characteristic clinical features support however the idea that these antibodies may somehow influence the presentation and course of the disease and possibly predict their response to therapy.

The retrospective nature of this study represents the major limitation of our work, especially in the analysis of the therapeutic response, altogether with the limited number of seropositive patients, the lack of complete antibody characterization of all included patients, and the lack of multiple testing correction in the statistical analysis. Our findings are nevertheless coherent with previous studies. The strengths of this study include the use of a national multicenter database collecting a consistent case series reflective of the Italian CIDP population avoiding case selection related to a previous lack of response to therapy or an acute onset of CIDP and the use of well-defined and uniform inclusion and assessment criteria. This lack of selection might explain the lower frequency of these antibodies compared to a previous Italian study where some of our patients had been also included [10].

In any instance, our study supports the fact that the implementation of a pathogenetic-oriented approach in the evaluation and diagnosis of patients with CIDP may help in the identification of patients with a peculiar clinical and immunological phenotype requiring different treatment strategies and theoretically support the hypothesis that CIDP might

be a syndrome that includes different chronic demyelinating neuropathies [33].

Author contribution Giuseppe Liberatore and Alberto De Lorenzo contributed to the conception of the research project, reviewed and commented on the statistical analysis, wrote the first draft of the report, and reviewed the report. Eduardo Nobile-Orazio conceived, organized, and designed the study; reviewed and commented on the statistical analysis; wrote the first draft of the report; and reviewed the report. Claudia Giannotta and Luis Querol contributed to perform laboratory analysis and reviewed the report. All the other authors contributed to the organization and execution of the research project, and reviewed and commented the report.

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Data availability Anonymized data will be shared by request from any qualified investigator upon reasonable request.

Declarations

Ethical approval The study was approved by the Ethical Committee of each participating center. All patients gave written informed consent.

Conflict of interest Dr. Eduardo Nobile-Orazio reported personal fees for advisory or scientific board from Kedrion, Italy; Baxter, Italy; Novartis, Switzerland; CSL Behring, Italy; LFB, France; and Astellas, the Netherlands, outside the submitted work and travel grants to attend scientific meeting from Baxter, Grifols, Kedrion, and Novartis, Italy. Dr. Pietro Emiliano Doneddu reported travel grants to attend scientific meetings from CSL Behring and Kedrion. Dr. Giuseppe Liberatore reported travel grants to attend scientific meetings from CSL Behring and Kedrion. Dr. Dario Cocito reported honoraria for lecturing from Shire, CSL Behring, and Kedrion and travel grants to attend scientific meeting from Shire, Kedrion, and CSL Behring. Dr. Erdita Peci reported travel grants to attend scientific meetings from CSL Behring. Dr. Raffaella Fazio has served on scientific advisory boards for CSL Behring and has received travel grants from Kedrion and CSL Behring to attend scientific meeting. Dr. Marinella Carpo reported travel grants to attend scientific meetings from Kedrion. Dr. Chiara Briani has served on scientific advisory boards for Pfizer and has received travel grants from Kedrion and CSL Behring to attend scientific meeting. Dr. Giuseppe Cosentino reported travel grants to attend scientific meetings from CSL Behring and Kedrion. Dr. Andrea Cortese reported travel grants to attend scientific meetings from Kedrion. Dr. Fiore Manganello reported personal fees for scientific events from CSL Behring and has received travel grants to attend scientific meetings from CSL Behring and Kedrion. Dr. Guido Cavaletti reported honoraria for lecturing and travel grants to attend scientific meetings from Kedrion. Dr. Massimiliano Filosto has served on scientific advisory boards for CSL Behring and Sarepta Therapeutics and has received travel grants from Sanofi-Genzyme, Kedrion, Baxter, and CSL Behring to attend scientific meeting. Dr. Girolama Alessandra Marfia reported consultancy fees and travel fundings from CSL Behring, Kedrion, Shire, and Grifols. Dr. Giovanni Antonini reported honoraria for lecturing from

Kedrion and Sanofi-Genzyme, and travel grants from Kedrion, Sanofi-Genzyme, and LJ Pharma. All the other authors declare no competing interests.

References


- Broers MC, Bunschoten C, Nieboer D, Lingsma HF, Jacobs BC (2019) Incidence and prevalence of chronic inflammatory demyelinating polyradiculoneuropathy: a systematic review and meta-analysis. *Neuroepidemiology*. 52(3-4):161–172. <https://doi.org/10.1159/000494291>
- Mathey EK, Park SB, Hughes RAC et al (2015) Chronic inflammatory demyelinating polyradiculoneuropathy: from pathology to phenotype. *J Neurol Neurosurg Psychiatry* 86(9):973–985. <https://doi.org/10.1136/jnnp-2014-309697>
- Doneddu PE, Nobile-Orazio E (2018) Management of chronic inflammatory demyelinating polyradiculopathy. *Curr Opin Neurol* 31(5):511–516. <https://doi.org/10.1097/WCO.0000000000000595>
- Doneddu PE, Cocito D, Manganelli F et al (2019) Atypical CIDP: diagnostic criteria, progression and treatment response. Data from the Italian CIDP Database. *J Neurol Neurosurg Psychiatry* 90(2):125–132. <https://doi.org/10.1136/jnnp-2018-318714>
- Kieseier BC, Mathey EK, Sommer C, Hartung H-P (2018) Immune-mediated neuropathies. *Nat Rev Dis Primer* 4(1):31. <https://doi.org/10.1038/s41572-018-0027-2>
- Ng JKM, Malotka J, Kawakami N et al (2012) Neurofascin as a target for autoantibodies in peripheral neuropathies. *Neurology*. 79(23):2241–2248. <https://doi.org/10.1212/WNL.0b013e31827689ad>
- Querol L, Nogales-Gadea G, Rojas-Garcia R et al (2014) Neurofascin IgG4 antibodies in CIDP associate with disabling tremor and poor response to IVIg. *Neurology*. 82(10):879–886. <https://doi.org/10.1212/WNL.0000000000000205>
- Ogata H, Yamasaki R, Hiwatashi A et al (2015) Characterization of IgG4 anti-neurofascin 155 antibody-positive polyneuropathy. *Ann Clin Transl Neurol* 2(10):960–971. <https://doi.org/10.1002/acn3.248>
- Devaux JJ, Miura Y, Fukami Y et al (2016) Neurofascin-155 IgG4 in chronic inflammatory demyelinating polyneuropathy. *Neurology*. 86(9):800–807. <https://doi.org/10.1212/WNL.00000000000002418>
- Cortese A, Lombardi R, Briani C et al (2020) Antibodies to neurofascin, contactin-1, and contactin-associated protein 1 in CIDP: Clinical relevance of IgG isotype. *Neurol - Neuroimmunol Neuroinflamm* 7(1):e639. <https://doi.org/10.1212/NXI.00000000000000639>
- Kadoya M, Kaida K, Koike H et al (2016) IgG4 anti-neurofascin155 antibodies in chronic inflammatory demyelinating polyradiculoneuropathy: Clinical significance and diagnostic utility of a conventional assay. *J Neuroimmunol* 301:16–22. <https://doi.org/10.1016/j.jneuroim.2016.10.013>
- Mathey EK, Garg N, Park SB et al (2017) Autoantibody responses to nodal and paranodal antigens in chronic inflammatory neuropathies. *J Neuroimmunol* 309:41–46. <https://doi.org/10.1016/j.jneuroim.2017.05.002>
- Delmont E, Manso C, Querol L et al (2017) Autoantibodies to nodal isoforms of neurofascin in chronic inflammatory demyelinating polyneuropathy. *Brain*. 140(7):1851–1858. <https://doi.org/10.1093/brain/awx124>
- Burnor E, Yang L, Zhou H et al (2018) Neurofascin antibodies in autoimmune, genetic, and idiopathic neuropathies. *Neurology*. 90(1):e31–e38. <https://doi.org/10.1212/WNL.0000000000004773>
- Querol L, Nogales-Gadea G, Rojas-Garcia R et al (2013) Antibodies to contactin-1 in chronic inflammatory demyelinating polyneuropathy: contactin-1 in Aggressive CIDP. *Ann Neurol* 73(3):370–380. <https://doi.org/10.1002/ana.23794>
- Miura Y, Devaux JJ, Fukami Y et al (2015) Contactin 1 IgG4 associates to chronic inflammatory demyelinating polyneuropathy with sensory ataxia. *Brain*. 138(6):1484–1491. <https://doi.org/10.1093/brain/awv054>
- Doppler K, Appeltshauer L, Wilhelmi K et al (2015) Destruction of paranodal architecture in inflammatory neuropathy with anti-contactin-1 autoantibodies. *J Neurol Neurosurg Psychiatry* 86(7):720–728. <https://doi.org/10.1136/jnnp-2014-309916>
- Doppler K, Appeltshauer L, Villmann C et al (2016) Auto-antibodies to contactin-associated protein 1 (Caspr) in two patients with painful inflammatory neuropathy. *Brain*. 139(10):2617–2630. <https://doi.org/10.1093/brain/aww189>
- Klehmet J, Märtschen S, Ruprecht K et al (2018) Analysis of anti-ganglioside antibodies by a line immunoassay in patients with chronic-inflammatory demyelinating polyneuropathies (CIDP). *Clin Chem Lab Med CCLM* 56(6):919–926. <https://doi.org/10.1515/cclm-2017-0792>
- Martinez-Thompson JM, Snyder MR, Ettore M et al (2018) Composite ganglioside autoantibodies and immune treatment response in MMN and MADSAM. *Muscle Nerve* 57(6):1000–1005. <https://doi.org/10.1002/mus.26051>
- Querol L, Siles AM, Alba-Rovira R et al (2017) Antibodies against peripheral nerve antigens in chronic inflammatory demyelinating polyradiculoneuropathy. *Sci Rep* 7(1):14411. <https://doi.org/10.1038/s41598-017-14853-4>
- Liberatore G, Manganelli F, Doneddu PE et al (2020) Chronic inflammatory demyelinating polyradiculoneuropathy: can a diagnosis be made in patients not fulfilling electrodiagnostic criteria? *Eur J Neurol*. Published online October 15, ene.14545. <https://doi.org/10.1111/ene.14545>
- Joint Task Force of the EFNS and the PNS (2010) European Federation of Neurological Societies/Peripheral Nerve Society Guideline on management of chronic inflammatory demyelinating polyradiculoneuropathy: report of a joint task force of the European Federation of Neurological Societies and the Peripheral Nerve Society - First Revision. *J Peripher Nerv Syst* 15(1):1–9. <https://doi.org/10.1111/j.1529-8027.2010.00245.x>
- Liberatore G, Giannotta C, Sajeev BP et al (2020) Sensitivity and specificity of a commercial ELISA test for anti-MAG antibodies in patients with neuropathy. *J Neuroimmunol* 345:577288. <https://doi.org/10.1016/j.jneuroim.2020.577288>
- Nobile-Orazio E, Terenghi F, Giannotta C, Gallia F, Nozza A (2009) Serum VEGF levels in POEMS syndrome and in immune-mediated neuropathies. *Neurology*. 72(11):1024–1026. <https://doi.org/10.1212/01.wnl.0000344569.13496.ff>
- Nobile-Orazio E, Gallia F, Terenghi F, Allaria S, Giannotta C, Carpo M (2008) How useful are anti-neural IgM antibodies in the diagnosis of chronic immune-mediated neuropathies? *J Neurol Sci* 266(1-2):156–163. <https://doi.org/10.1016/j.jns.2007.09.020>
- Hughes RAC, Bensa S, Willison HJ et al (2001) Randomized controlled trial of intravenous immunoglobulin versus oral prednisolone in chronic inflammatory demyelinating polyradiculoneuropathy. *Ann Neurol* 50:195–201. <https://doi.org/10.1002/ana.1088>
- Breiner A, Bourque PR, Allen JA (2019) Updated cerebrospinal fluid total protein reference values improve chronic inflammatory demyelinating polyneuropathy diagnosis. *Muscle Nerve* 60(2):180–183. <https://doi.org/10.1002/mus.26488>
- Zhang X, Zheng P, Devaux JJ et al (2019) Chronic inflammatory demyelinating polyneuropathy with anti-NF155 IgG4 in China. *J Neuroimmunol* 337:577074. <https://doi.org/10.1016/j.jneuroim.2019.577074>

30. Delmont E, Brodovitch A, Kouton L et al (2020) Antibodies against the node of Ranvier: a real-life evaluation of incidence, clinical features and response to treatment based on a prospective analysis of 1500 sera. *J Neurol* 267(12):3664–3672. <https://doi.org/10.1007/s00415-020-10041-z>
31. Cortese A, Devaux JJ, Zardini E et al (2016) Neurofascin-155 as a putative antigen in combined central and peripheral demyelination. *Neurol Neuroimmunol Neuroinflamm* 3(4):e238. <https://doi.org/10.1212/NXI.0000000000000238>
32. Kuwahara M, Suzuki H, Samukawa M, Hamada Y, Takada K, Kusunoki S (2013) Clinical features of CIDP with LM1-associated antibodies. *J Neurol Neurosurg Psychiatry* 84(5):573–575. <https://doi.org/10.1136/jnnp-2012-303440>
33. Nobile-Orazio E (2014) Chronic inflammatory demyelinating polyradiculoneuropathy and variants: where we are and where we should go. *J Peripher Nerv Syst* 19(1):2–13. <https://doi.org/10.1111/jns5.12053>

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