Raisins and Currants as Conventional Nutraceuticals in Italian Market: Natural Occurrence of Ochratoxin A

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Abstract: The healthy consumers make a strong pressure to natural products that can prevent the chronic diseases and improve the general health status, and therefore an important aspect that have to be considered is the safe level of the nutraceuticals. This study reports the occurrence of Ochratoxin A (OTA) and associated fungal contamination in 35 samples of dried vine fruits imported in the European community potentially used for the development of new nutraceutical supplements. High pressure liquid chromatography analysis identified 18 samples as contaminated by OTA with an average level of 2.6 μ g/kg. OTA was measured in 4 samples of currants (mean value of 6.6 μ g/kg) and 13 samples of raisins (mean value of 1.4 μ g/kg). In one sample of currants and one of raisins from Turkey OTA exceeded the limits set by European Commission of 10 μ g/kg, being contaminated with 12.61 and 15.99 μ g/kg, respectively. All the positive samples were confirmed by Orbitrap Q Exactive through their molecular weight and the corresponding fragmentation. The worldwide consumption of dried vine fruits contributed to OTA exposure in several group of consumers. In particular, considering the potential nutraceutical approach, this consumption may be represent a severe risk for healthy consumers that consider these products like healthy and salutistic for their contents in antioxidants, flavonoids, and polyphenols. Data reported in this study confirmed the need to regularly monitor mycotoxin levels in these food products and optimize the process of fruits drying in order to reduce the development of toxigenic molds.

Keywords: *Aspergillus* spp., dried vine fruits, food safety, nutraceutical food, ochratoxin A, supplement

Introduction

Food Chemistry

In the last few years, consumers have shown a significant interest to include a high level of bioactive compounds in their standard diet, preferably derivated from natural sources such as plants and fruits (Bhat and others 2010), encouraging the development of new nutraceutical foods and supplements by researchers and pharmaceutical companies. The term nutraceutical was coined by DeFelice (1995), referring to any substance that may be considered a food or part of a food providing medical or health benefits. More recently, the Dietary Supplement Health and Education Act defines nutraceuticals as a dietary supplement that may contain an herb or other botanical, or a concentrate, metabolite, constituent, extract, or combination of any ingredient from the other categories (Frankos and others 2010). In this frame, numerous studies have strongly suggested that the inclusion of grapes and grape products as supplements incorporated in daily intake of foods may generate significant health benefits, mainly due to the presence of phenolic compounds, that are potent antioxidants recognized as preventative agents against several diseases and protecting the body tissues against oxidative stress (Shalaby and Horwitz 2015). Global raisin production for 2014/2015 is forecast at a record 1.2 million metric tons, up 5% from the previous year (USDA 2014). Consumption, which lagged production of the previous 4 years, is also at a record 1.2 million tons. Dried vine fruits are produced by dehydrating grape berries which is of particular interest because bioactive molecules must be ingested in a strict dose-dependent manner to achieve beneficial therapeutic.

On the other side, raisins and currants are often affected by toxigenic fungal colonization, in particular *Aspergillus* section *Nigri* or black aspergilli, with higher occurrence of *Aspergillus niger* aggregate (Perrone and others 2007; Palumbo and others 2011), which are known to produce Ochratoxin A (OTA). This is a nephrotic mycotoxin classified by the International Agency for Research on Cancer (IARC) as possibly carcinogenic to humans (group 2B; IARC 1993). OTA was evaluated by the Scientific Committee on Food (SCF) in 1998 when it was concluded that it has carcinogenic, nephrotoxic, teratogenic, immunotoxic, and possibly neurotoxic properties (SCF 1998).

The presence of OTA has been widely reported in dried vine fruits (MacDonald and others 1999; Ostry and others 2002; Möller and Nyberg 2003; Stefanaki and others 2003) in many countries (Lombaert and others 2004; Covarelli and others 2012) and with different levels, usually much higher than wine, as reported in Europe by Miraglia and Brera (2002). Dried vine fruits from Athens and Thebes market were analyzed; 23% of samples were found to be contaminated at high levels and 18 exceeded the EU limit (Kollia and others 2013). Samples purchased from Tunisian and Spanish markets were analyzed for the presence of 16 mycotoxins and revealed the contamination with 2 to 6 mycotoxins in 51% of the analyzed samples (Azaiez and others 2015).

These evidences confirmed the concern related to the consumption of these products, which contributed to a significant extent to OTA exposure, especially in vulnerable groups of

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consumers such as children (Raiola and others 2015). For this reason, European Commission (EC) established the limit of 10.0 μ g/kg for OTA in dried vine fruit (EC 2006).

The health hazard of nutraceutical formulations potentially obtained from dried vine fruits products largely depends on the presence of unusually high concentrations of toxic compounds. More purposely, if these products are used for infusion preparations or extracted and dehydrated from pharmaceutical companies in order to concentrate the antioxidant compounds, also an higher level of undesirable compounds such as mycotoxins would be available for consumers (Kalny and others 2007).

In addition, a significant increase of health risk in contaminated supplements could derive from the different physical state compared with the food source. In fact, the matrix in which a contaminant is present plays an important role in its bioaccessibility and bioavailability.

For these multiple reasons, a particular attention should be addressed to the control quality of the foodstuffs used as a source of nutraceutical supplements. However, an important limit for the safety of nutraceutical preparations is represented by the absence of a formal legislation regulating these products (Gulati and Ottaway 2006; Martínez-Domínguez and others 2014).

The first goal of this study was to evaluate the incidence of fungal and yeast contamination in raisins and currants of different origins intended to use as raw materials for the development of nutraceutical formulations in Italian market. Successively, we investigated the OTA occurrence in the same products by an innovative and optimized method developed in our laboratory, based on the confirm of positive samples by UHPLC-HRMS Orbitrap Q Exactive.

Materials and Methods

Samples

A total of 35 dried vine fruit samples from worldwide origin and the packaged samples were obtained from an importing company from Turkey (Fundacio Privada International Tree Nut, Antalya) or purchased from different markets in Bari (Italy). Samples were transported to laboratory and stored in air-tight containers at room temperature until mycological and chemical analysis.

Mycological analysis

Mycological analysis of dried vine fruits was performed by direct plating onto Dichloran Rose Bengal Chloramphenicol (DRBC, Oxoid) agar (King and others 1979; Pitt and Hocking 1997). Representative monoconidial isolates, based on diversity and incidence within each sample, were finally stored in cryotubes From each sample, 50 dried berries were aseptically plated (5 for plate) in DRBC agar petri dishes (90 mm).

Plates were incubated in darkness for 7 days at 25 °C and examined daily. After incubation, the number of berries showing fungal infection were recorded and the incidence of fungal genus for each sample was determined. Fungal colonies were subcultured both on malt extract agar (MEA) and Czapek–Dox Agar containing 100 mg/L of streptomycin and 50 mg/L of neomycin sulphate as antibacterial agents.

The strains were identified to genus/section level by culture and morphological observation under stereoscope (Pitt 1979; Samson and others 2004). Representative monoconidial isolates from each sample were finally stored in cryotubes under 20% glycerol at −80 °C and under liquid nitrogen for successive molecular characterization.

Chemical and materials

OTA stock solution (1 mg/mL) was prepared by dissolving the solid standard purchased from Sigma Aldrich (Milan, Italy) in methanol (99:1, v/v). Standard solutions for HPLC calibration or spiking purposes were prepared by diluting stock solution. Acetonitrile, methanol, water (HPLC grade), and glacial acetic acid (CH3COOH) were purchased from Mallinckrodt Baker (Milan, Italy).

Potassium chloride (KCl), monosodium phosphate (NaH_2PO_4) , sodium sulphate $(NaSO_4)$, sodium chloride (NaCl), and sodium bicarbonate (Na $HCO₃$) were obtained from Sigma Aldrich.

Acetonitrile, methanol, and water for UHPLC mobile phase and organic solvents were LC/MS grade from Merck (Darmstadt, Germany), whereas formic acid and ammonium formate were obtained from Fluka (Milan, Italy).

Sample preparation

The samples were extracted and purified according to Vicam (1997) procedure for OTA in dried vine. As regards the retail unpacked dried vine fruits, 1 kg was purchased, from which 5 incremental samples weighing 100 g each were taken; consequently, the laboratory samples were 500 g that were homogenized in a Waring blender. Analytical method was similar to that described by Karami-Osboo and others (2015) with slight modifications.

Fifty grams of homogenized sample was used for extraction with 100 mL methanol containing 1% sodium bicarbonate solution (70:30) in Polytron (Lucerne, Switzerland). The mixture was homogenized for 1 min and filtered with filter paper. An aliquot of 10 mL was diluted with 40 mL phosphate-buffered saline (0.2 g KH_2PO_4 , 1.1 g anhydrous Na₂HPO₄, 8.0 g NaCl, 0.20 g KCl, pH 7.0) and filtered. An aliquot of 10 mL was passed through the Ochratest TM (Ochratest, VICAM, Watertown, Mass., U.S.A.) immunoaffinity column (2 drops/s), then the column was washed with 20 mL of water and eluted with 4 mL of methanol. The methanolic extract was dried with a centrifugal evaporator, resuspended in 1 mL of methanol, filtered with a 0.22μ m filter (Phenomenex, Torrance, Calif., U.S.A.) and injected in the LC apparatus. Each sample was analyzed in triplicate.

Recovery experiments and linearity range

Recovery tests were carried out dried vine in triplicate. OTAfree samples were spiked with OTA standard (Sigma) and maintained at room temperature in a dark room overnight. Four contamination levels used were: 1, 10, 50, and 100 μ g/kg. Linearity was estimated in the working range of 0, 1 to 50 ng/mL for OTA, with 4 calibration levels, each injected in duplicate.

HPLC analysis

OTA analysis was performed using an LC (Shimadzu-Japan) equipped with an autosampler SIL-20A, 2 pumps LC-20AD and a fluorimetric detection RF- 20AXL (OTA: $\lambda_{\rm ex} = 360$ nm; $\lambda_{\rm em} = 460$ nm). An Onyx Monolithic column with 3 μ m particle size C_{18} (100 × 3.0 mm; Phenomenex) was employed. Mobile phase was used in isocratic conditions: 65% A (1% CH₃COOH in H_2O), 35% B (1% CH₃COOH in CH₃CN) at the flow rate of 1 mL/min (Raiola and others 2012).

OTA confirmation by Orbitrap Q Exactive analysis

Positive samples for the presence of OTA found by HPLC were confirmed by the high-resolution mass spectrometry

Table 1–Recovery ± RSD (%), LOD and LOQ (ng/g).

OTA level $(\mu$ g/kg)	Recovery \pm $RSD(\%)$	LOD (ng/g)	LOQ (ng/g)
	85.2 ± 2.3	0.05	0.15
10	88.1 ± 5.1	0.05	0.15
50	84.6 ± 3.5	0.05	0.15
100	89.0 ± 5.7	0.05	0.15

(HRMS-Orbitrap) analysis, by using an optimized and innovative method carried out in our laboratory.

Qualitative profile of OTA has been obtained using ultra high pressure liquid chromatography (UHPLC, Thermo Fisher Scientific, Waltham, Mass., U.S.A.) equipped with a degassing system, a Dionex Ultimate 3000 a Quaternary UHPLC pump working at 1250 bar, an auto sampler device and a thermostated $(T =$ ⁵⁰ °C) Thermo fisher AccucoreAq C18 (100 [×] 2.1 mm) column 2.6 μ m particle size. Injection volume was of 5 μ L. Eluent phase was formed as follow: phase A $(H₂O$ in 0.1% formic acid and 5 mM ammonium formate), phase B (methanol in 0.1% formic acid and 5 mM ammonium formate). OTA has been eluted using a 0.4 mL/min flow rate with a gradient programmed as follows: 0 min—60 % of phase B, 9 min—100% of phase B, 12 min—100% of phase B, 12.1 min—60% of phase B, 15 min—60% of phase B.

For the mass spectrometry analysis a Q Exactive Orbitrap LC-MS/MS (Thermo Fisher Scientific) has been used. An ESI source (HESI II, Thermo Fischer Scientific) operating inpositive (ESI+) ion mode for OTA analysis. Ion source parameters in ESI+ mode were: spray voltage 3.50 kV, sheath gas $(N_2 > 95%)$ 30, auxiliary gas ($N_2 > 95\%$) 10, capillary temperature 320 °C, S-lens RF level 50, auxiliary gas heater temperature 300 °C.

Value for automatic gain control (AGC) target was set at $1 \times$ 106, with a resolution of 70000 full width at half maximum (FWHM), and a scan rate in the range between 90 and 1000 *m*/*z*. The experiment was performed in full MS scan.

The accuracy and calibration of the Q ExactiveOrbitrap LC-MS/MS was checked daily using a reference standard mixture obtained from Thermo Fisher Scientific. Data analysis and processing has been performed using the Xcalibur software, v. 3.0.63 (Xcalibur, Thermo Fisher Scientific).

OTA was identified by its retention time and exact mass at 5 decimal places from the total ion chromatogram (TIC) (Figure 1).

Results

Recovery and LOD

Table 1 showed values for recovery, the detection limit (LOD), and limit of quantification (LOQ) of OTA obtained by HPLC analysis. Values of recovery were 85.2 \pm 2.3%, 88.1 \pm 5.1%, 84.6 \pm 3.5%, and 89.01 \pm 5.7% for the levels of contamination of 1, 10, 50, and 100 μ g/kg, respectively. LOD obtained was 0.05 ng/g for OTA whereas LOQ was 0.15 ng/g. Although interday precision was calculated by the relative standard deviation (RSDR) calculated from concentration results of spiked samples generated under reproducibility conditions by one determination per concentration on 6 d, RSD_R for the procedures validated at LQ and 10 LQ were lower than 12% and 8%, respectively. These results demonstrated that the developed methods yielded acceptable recoveries from the analyzed samples.

Fungal contamination in dried vine fruits

Table 2 reported the incidence of contamination by *Aspergillus sect. Nigri* spp. and other species in dried vine fruits samples. The global incidence of contamination by *Aspergillus sect. Nigri* resulted very high. In 19 over 35 samples 100% of the fruits resulted contaminated by fungi belonging to this section. With respect to the type of dried fruits, although in 10/17 raisins sample the contamination was 100%, the average incidence of *Aspergillus* section *Nigri* was slightly higher on currants (74%) than on raisins (72%). With respect to the origin, samples from Turkey were the most contaminated by *Aspergillus sect. Nigri* (range of incidence between 86% and 100%), with an average occurrence of 94%. Eighteen samples showed 100% of contamination, whereas in only 1 sample out of 21 no *Aspergillus* occurred. *Penicillium* spp. were isolated in 4 samples, from Iran, Chile, South Africa, and the U.S., whereas *Erotium* spp. were found in 2 samples from Chile, with an incidence of 58% and 44% (Table 2).

In the only sample from Turkey where no *Aspergillus* was detected, we found a high incidence of *Monascus* spp. (46%; Table 2). In the samples from Chile, *Aspergillus* incidence was in the range

Figure 1–Orbitrap Q Exactive spectrum of OTA.

between 12% and 72%, whereas 2 of 7 samples resulted not contaminated by any fungal species. In 2 samples of currants from Iran, levels of *Aspergillus* incidence equal to 8% and 54% were observed, whereas in one sample *Penicillium* spp. were isolated.

In 2 samples of green raisins from China, *Aspergillus sect. Nigri* incidence was between 82% and 100%, whereas in 2 raisins samples from the U.S., the levels were 42% and 44%. The sample of golden raisins from South Africa was not contaminated by *Aspergillus* but by *Penicillium* spp., however with a low incidence of 2% (Table 2).

OTA contamination in dried fruits

Table 3 shows the incidence of OTA contamination in dried vine fruits samples from different geographical origin. The presence of OTA in positive fruits found by HPLC was confirmed by Orbitrap Q Exactive analysis.

With respect to the type of dried fruits, the relative occurrence of contaminated samples in raisins (14/22) was higher than in currants (4/13). Nevertheless, in the latter, the average level of contamination was higher than in raisins (4.1 μ g/kg versus 1.35 μ g/ kg). Levels of OTA ranged from 0.05 μ g/kg to 15.99 μ g/kg in currants and from 0.95 μ g/kg to 12.61 μ g/kg in raisins.

With respect to geographical origin, the higher average contamination was measured in samples from Turkey (3.1 μ g/kg). In 65% of samples from this state OTA levels were between 0.05 and 15.99 μ g/kg. In 2 samples levels of OTA over the European Union (EU) legal limit of 10 μ g/kg were measured (15.99 and 12.61 μ g/kg in currants and raisins respectively).

OTA occurred in only 2 above 7 samples from Chile, with the values of 1.09 and 1.18 μ g/kg for golden raisins and raisins, respectively. In both 2 samples of currants from Iran, as in the samples from China, although a high fungal contamination was observed, OTA was not detected.

In 2 samples of raisins from the U.S., OTA was measured at levels equal to 1.07 and 1.14 μ g/kg. The sample of golden raisins from South Africa was contaminated by OTA in the amount of 1.91 μ g/kg.

Discussion

This study examined an important problematic in the context of food safety that is the quality control of the raisins and currants, potentially adopted for the development of nutraceutical supplements, because it is widely reported their healthy effect, attributable to the high amount of polyphenols, including flavonoids, anthocyanins, proanthocyanins, and stilbenes.

In particular, we evaluated the incidence of fungal contamination and OTA amount originating from several countries of the world by an innovative and optimized method of detection.

We found low levels of OTA contamination in sample of dried vine fruits imported from countries outside Europe, despite the high incidence of potential toxigenic fungi occurring in the samples. These data are in agreement with previous studies performed in different countries and regions, which described dried vine fruits as susceptible of fungal growth and subsequent mycotoxin

Table 3–OTA contamination in dried vine fruits.

Sample	Dried vine fruits	Origin	OTA $(\mu g/kg)$
DF ₁	Currants	Turkey	n.d.
DF ₂	Currants	Chile	n.d.
DF19	Currants	Turkey	n.d.
DF ₂₅	Currants	Turkey	n.d.
DF26	Raisins	Chile	n.d.
DF41	Currants	Iran	n.d.
DF52	Currants	Turkey	n.d.
DF53	Raisins	USA	1.14
DF78	Currants	Turkey	n.d.
DF79	Currants	Turkey	n.d.
DF80	Raisins	Chile	n.d.
DF81	Currants	Iran	n.d.
DF82	Currants	Turkey	0.23
DF83	Currants	Turkey	0.13
DF91	Green raisins	China	n d-
DF95	Raisins	Chile	n.d.
DF96	Raisins	Chile	n.d.
DF97	Currants	Turkey	15.99
DF105	Currants	Turkey	0.05
DF106	Raisins	Turkey	12.61
DF107	Raisins	Turkey	1.18
DF108	Raisins	Turkey	n.d.
DF109	Raisins	Turkey	1.3
DF110	Raisins	Turkey	2.17
DF111	Raisins	Turkey	n.d.
DF112	Raisins	Turkey	0.95
DF113	Raisins	Turkey	1.41
DF114	Raisins	Turkey	1.54
DF115	Raisins	Turkey	1.05
DF120	Golden raisins	South Africa	1.9
DF121	Raisins	Chile	1.18
DF122	Raisins	USA	1.07
DF123	Golden raisins	Chile	1.09
DF124	Sultana	Turkey	1.12
DF125	Green raisins	China	n.d.

n.d., not detected

production and accumulation. In 2002, Miraglia and Brera summarized data on dried fruits contamination provided by European countries, indicating these commodities to be highly exposed to OTA contamination. The incidence reported for dried vine fruits was 73% in Europe (over 800 samples analyzed) with OTA levels around 3 μ g/kg.

Stefanaki and others (2003) investigated the presence of OTA in 81 samples of Greek domestic currants and sultanas. The latter resulted less contaminated than currants (average level of 2.1 and 2.8 μ g/kg, respectively), and globally 7.5% of both types of dried vine fruit exceeded OTA limit of 10 μ g/kg. In Italy, Imperato and others (2011) defined dried vine fruits as the main food matrix contaminated by OTA within the survey carried on 345 samples of imported foodstuffs. However, the contamination levels were below the maximum tolerable limit set by EU regulation.

Zinedine and others (2007) analyzed 20 samples of dried raisins purchased from retail shops in Morocco. OTA amounts ranged from 0.05 to 4.95 μ g/kg in dried raisins in the samples.

Investigations of dried vine fruits samples from Turkey showed the presence of samples which overtook the EU limits: Aksoy and others (2007) reported OTA mean value of 1.36 μ g/kg, whereas Bircan (2009) measured detectable levels in the range from 0.5 to 58 μ g/kg in 53% of the samples.

It is important to note that, despite the high occurrence of *Aspergillus sect. Nigri*, potentially able to produce high amount of OTA, the average level of OTA measured in the positive samples was relatively low (2.56 μ g/kg). However, 2 samples, 1 raisins and

1 currants, both from Turkey, exceeded EU legal limit with 12.61 and 15.99 μ g/kg, respectively.

Biodiversity of fungal species isolated from our samples is consistent with contamination usually reported on dried vine fruits (Abarca and others 2003; Magnoli and others 2004; Romero and others 2005; Tournas and others 2015). Besides *Aspergillus* and *Penicillium* species, we also detected *Eurotium*, *Monascus*, and other yeasts. Interestingly, in one sample from Turkey (Table 2) only *Monascus* strains occurred. Organisms of the genus *Monascus*, known for the production of red pigments used as coloring agents in food and textiles (Fabre and others 1993), are able to grow over several substrates and food by-products, including dried fruits (Water Relations of Foods 1974; Golob and others 2002).

Results presented in this paper revealed a significant variance of the incidence of *Aspergillus* and OTA development. On the contrary, OTA occurrence is not correlated with the incidence of fungal contamination. This discrepancy may be due to the specific environmental conditions developed by processing grape berries. During ripening of the fruits (Covarelli and others 2012) temperature increases and the sugar is concentrated as the moisture content decreases. By this way drying fruits become an optimal and selective medium for xerotolerant molds such as *A. niger sect. Nigri* species. However, these conditions are not necessarily optimal for mycotoxin production as several studies confirmed (Leong and others 2006; Magan 2007). Recently, Passamani and others (2014) demonstrated that the best range for OTA production by *A. niger* and *A. carbonarius* was at 15 °C, aw 0.99, whereas the growth was optimal at 20 to 30 °C and 24 to 37 °C, respectively. Zhang and others (2014) analyzed a total of 56 dried vine fruits from Chinese markets. All samples were analyzed for OTA. Only one sample exceeded the EU. Heshmati and others (2015) evaluated the occurrence of OTA in 66 samples of dried grapes in Iran. OTA was detected in 57.5% of currants, 62.5% of sultanas, and 60% of raisins samples. None of the samples contained OTA level exceeding the maximum limit prescribed in the EU regulations, which is 10 μ g/kg.

In general, to prevent or reduce the occurrence of species of molds able to produce OTA in dried fruits, good agricultural practices, good manufacturing practices, and hazard risk analysis should be applied. As an example, the use of drying chambers instead of sun drying could significantly reduce the fungal development and the corresponding OTA production.

However, an interesting aspect to consider in the frame of OTA occurrence in nutraceutical products is the potential protective action exercised by some antioxidants, such as α -tocopherol, cyanidin-3-o- β glucopyranoside ascorbic acid, and polyphenols against OTA effects on health.

These properties are probably due to the ability that the antioxidants have to act as superoxide anion scavengers, protecting cell membrane from the mycotoxin-induced damage (Atroshi and others, 2002). *In vitro* and *in vivo* studies (Baldi and others 2004; Costa and others 2007) showed that these substances could counteract some of the OTA toxic effects.

According to our study OTA occurrence is not at level of concern, except for 2 samples from Turkey. Despite these reassuring results, it should be considered that the low OTA levels found in analyzed samples, could represent a very significant exposure when these products are extracted to obtain nutraceutical supplements, where the level of phytochemicals, but also of contaminants are significantly concentrated, with the consequent risk to exceed the legislative limit fixed for OTA equal to 10 μ g/kg.

Conclusion

This study confirms the need to control mycotoxin levels with a regular monitoring activity, especially in area where the production of dried vine fruits is widely practiced.

In addition, because healthy consumers tend to join and to follow a better life style and to insert in the diet the food integrators or nutraceuticals to improve health status, more rigorous controls over the safety and quality of these products could be achieved through good manufacturing practice, regulatory control, and research efforts (Kho and Woo 2000). Although storage is generally identified as the most critical stage for OTA contamination, control strategies applied during the preharvest time result effective to reduce fungal infection (Amézqueta and others 2009).

Regulators should also employ actions to minimize these risks, by introducing specific limits for mycotoxins and in particular OTA for this particular category of products.

Our study revealed a significant variance of the incidence of *Aspergillus* and OTA development, whereas OTA occurrence is not correlated with the incidence of fungal contamination. This effect may be due to the specific environmental conditions developed by processing grape berries. The confirmation of OTA identification was carried out by an innovative method that is represented by Orbitrap Q Exactive, based on the molecular weight and the corresponding fragmentation. In one sample of currants and one of raisins from Turkey OTA exceeded the limits set by European Commission of 10 μ g/kg. Therefore, further investigations about the exposure to other mycotoxins from both conventional and innovative nutraceutical sources are need in order to obtain a complete risk assessment associated to their consumption. In addition, the impact of the different matrixes represented by dried vine fruits and nutraceutical supplements on bioaccessibility and bioavailability of OTA should be evaluated.

Acknowledgments

We thank Antonio Dimundo for his valuable technical assistance. This work was financially supported by EC KBBE-2007- 222690-2 MYCORED.

Conflict of Interest

The author(s) confirm that this article content has no conflict of interest.

Authors' Contributions

F. Fanelli and G. Cozzi carried out analyses of fungal contamination and ochratoxin A in dried vine fruits and wrote the manuscript; A. Raiola, I. Dini, and G. Mule analyzed data and ` wrote the manuscript; A.F. Logrieco and A. Ritieni designed the study, interpreted the results, and revised the manuscript.

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